

Journal section: Operative Dentistry and Endodontics  
Publication Types: Research

doi:10.4317/jced.53592  
<http://dx.doi.org/10.4317/jced.53592>

## Microhardness and shear bond-strength of carious dentin after fluorescence-aided or conventionally excavation: (An *in-vitro* comparison)

Uzay Koç-Vural, Esra Ergin, Sevil Gurgan

Hacettepe University, Department of Restorative Dentistry, 06100 Sıhhiye/Ankara

Correspondence:  
Hacettepe University  
Department of Restorative Dentistry  
06100 Sıhhiye/Ankara  
[uzaykoc@gmail.com](mailto:uzaykoc@gmail.com)

Koç-Vural U, Ergin E, Gurgan S. Microhardness and shear bond-strength of carious dentin after fluorescence-aided or conventionally excavation: (An *in-vitro* comparison). J Clin Exp Dent. 2018;10(7):e668-72.  
<http://www.medicinaoral.com/odo/volumenes/v10i7/jcedv10i7p668.pdf>

Received: 06/11/2016  
Accepted: 14/03/2018

Article Number: 53592 <http://www.medicinaoral.com/odo/indice.htm>  
© Medicina Oral S. L. C.I.F. B 96689336 - eISSN: 1989-5488  
eMail: [jced@jced.es](mailto:jced@jced.es)  
**Indexed in:**  
Pubmed  
Pubmed Central® (PMC)  
Scopus  
DOI® System

### Abstract

**Background:** To compare the fluorescence-aided and conventionally excavated dentin with microhardness and shear bond strength(SBS) tests.

**Material and Methods:** Twenty-four teeth with dentin caries were bisected through the center of the lesion into two halves. Forty-eight dentin specimens were embedded and mounted in an acrylic resin. All carious tissue was removed and classified as caries free using conventional visual tactile criteria. Then half of the specimens(n=24) were reinspected with fluorescence-aided caries excavation light(FACE) (FaceLight, W&H Dentalwerk, Bürmoos GmbH, Austria). Specimens were subjected to microhardness and shear bond strength testings. The fracture mode analysis was also performed. The data were compared with Student's t test and Chi-square test.

**Results:** Residual caries was observed in 2 out of 24 conventionally excavated specimens with FACE inspection( $p>0.05$ ). Mean Vickers hardness of the dentin was  $61.5\pm 5$  in the FACE group and  $70.3\pm 3$  kg/mm<sup>2</sup> in the conventionally excavated group( $p>0.05$ ). The mean SBS value of FACE group was  $11.42\pm 1.63$  MPa and  $18.27\pm 1.43$  MPa in conventionally excavated group. There was no statistically significant difference between conventional and FACE groups for microhardness and SBS tests( $p>0.05$ ). There were also no significant differences on the fracture mode distributions of the groups( $p>0.05$ ).

**Conclusions:** FACE method could be considered as a promising technique for removing infected dentin.

**Key words:** FACE, conventional excavation, residual caries detection, shear bond strength, microhardness.

### Introduction

Caries excavation has been still regarded as a confusing procedure for the dental practitioners. The concept of minimally invasive dentistry requires the selective removal of carious dentin which consists two layers (1). The outer infected layer, which is heavily infected by bacteria and irreversibly denatured has to be removed

while the only partially demineralized inner affected layer, which has the potential of remineralization, should be preserved during caries excavation process (2).

Many excavation methods have currently been used for the removal of infected dentin. In the conventional excavation technique, dentists determine the endpoint of the excavation by the hardness and color of the dentin

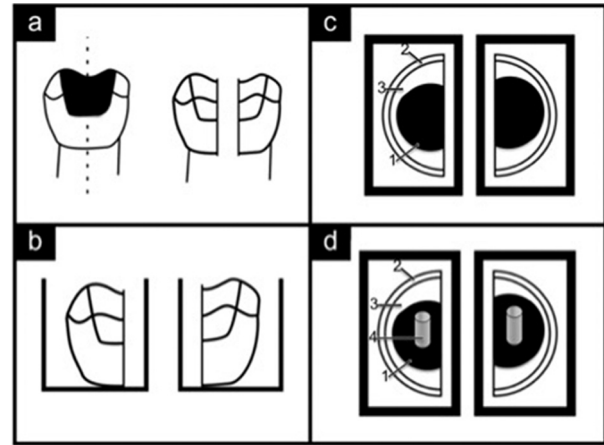
(3). Clinicians keep excavating carious dentin until reaching to the mineralized layer, considering the depth of excavation based on the dentin hardness and color. However, up to date, no certain criteria have been specified yet to discriminate the carious dentin layers. In the traditional visual-tactile inspection technique performed with a mirror and an explorer; it is considered that “the probe should not stick in the dentin and should not give a tug-back sensation” (4). However, the demineralized affected dentin, which can remineralize in time, could be sacrificed during the conventional excavation (5-7). Since this sensation is highly subjective, dye staining has been developed to make this procedure more objective by indicating only demineralized tissues (8,9). It has been previously reported that dye staining and bacterial penetration are different phenomena since dye staining might be a poor indicator for the discrimination of carious dentine layers (10). Therefore, alternative dentin caries removal methods using optical aids have been recently developed (11).

Fluorescence aided caries excavation (FACE), a novel caries excavation system has been claimed to be an objective method in the removal of infected dentin (12). In the FACE system, sound dentin fluoresces green after illuminated with violet light, whereas bacterially infected dentin emits red-orange fluorescence. When an exposed cavity is illuminated by the violet light, which causes the dental hard tissue to autofluorescence (wavelength of 405 nm), the porphyrins exuded by oral bacteria show a red fluorescence, indicating the essential areas for caries excavation (12,13). Dental hard tissues fluoresce green and carious dental hard tissues fluoresce orange-red under the FACE light. FACE has been shown to be more effective than conventional excavation in the removal of infected dentin *in vitro* (14-16).

Until now, little attention has been paid to the relation between hardness and the bond strength to dentin inspected with FACE. Thus, the aim of this study was to evaluate the microhardness and shear bond strength (SBS) of dentin, after inspected by FACE and to compare this technique with the visual-tactile method.

## Material and Methods

The experimental procedures were approved by the local ethics committee (GO 14/106). Freshly extracted, permanent maxillary or mandibular molars with occlusal caries lesions on the central fissure were collected and stored in 0.05 % formaldehyde solution at 4°C up to 1 month. Among those, the teeth having; a) caries lesions limited on enamel or reaching up to the pulp chamber, b) any restoration, c) sign of tetracycline staining and d) any cracks were excluded. Finally 24 teeth with visually similar sized caries lesions were selected and bisected through the center of the lesion in the occluso-apical direction to minimize the anatomical variations (Fig.



**Fig. 1:** Preparation of the specimens. 1-Caries excavated dentin, 2-Enamel, 3-Sound dentin, 4-Composite resin disc. (a) Teeth were bisected through the center of the lesion in the inciso-apical direction into the two halves. (b) The obtained sections were embedded in a chemically cured acrylic resin. (c) The occlusal surfaces were exposed (d) A resin composite disc was placed over dentin for SBS testing.

1a). The obtained sections were embedded in a chemically cured acrylic resin (Fig. 1b) (Vertex, Vertex Dental, Zeist The Netherlands), so that the occlusal surfaces could be exposed (Fig. 1c).

Then the specimens were divided into 2 groups representing similar lesion morphology (n=24). This allowed the allocation of the teeth into two groups of 24 teeth each, so that each group had the similar lesions before the excavation (Fig. 1c). The specimens were soaked in distilled water immediately at the dough stage of resin to avoid the harmful effect of temperature rise during polymerization, and were kept in distilled water at room temperature until hardness measurements were done within 24 h.

**Caries Excavation:** One operator performed the caries removal procedures (UKV) to minimize variations. Carious dentine of all specimens (n=48) was excavated conventionally. Soft dentin was removed by the help of round steel burs with different sizes without water-cooling in a low speed hand-piece and straight spoon excavators. The carious dentine was removed until the operator assessed the specimen as “free from caries” using the visual-tactile criteria. The preparation was finalized when the probe did not give a tug-back sensation, and clinically seemed sound. The hard dentin was preserved. Half of the conventionally excavated specimens, having similar lesions with the remaining 24 specimens, were inspected with FACE light (FaceLight, W&H Dentalwerk, Bürmoos GmbH, Austria) technique. This device generated fluorescence violet light (405 nm) by the help of 100-130-watt xenon discharge lamp to inspect the surfaces of the specimens (n=24). The room was darkened during the inspection procedure. Red-orange colored areas were further excavated until the transform was completed from orange-red to the green fluorescing area.

Hardness test: All hardness measurements were performed on the excavated surfaces of the specimens. An adhesive tape was adapted to the deepest cut surface of each specimen to ensure the excavated area and the surface was flattened by hand-polishing with 800,1200, 2500, and 4000 grid wet silicon carbide abrasive papers, respectively (Fig. 2). After the adhesive tape was removed, they were dried briefly using compressed air before the hardness test.

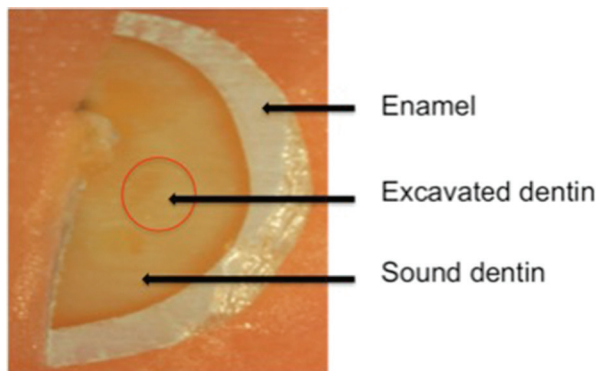


Fig. 2: A prepared specimen.

Vickers hardness tester (HMV, Shimadzu Corporation, Japan) was used to obtain precise hardness profiles of the excavated dentin surfaces. A load of 1 Newton (100 g) was applied for 15 s to produce the indentations. The Vickers hardness numbers (VHNs) were measured at five points in dentin with a minimum distance of 40 μm. Each line of indentations was placed in the area to ensure the presence of the excavated dentin.

Shear-bond strength (SBS) test: Following the hardness test, the excavated dentin surfaces were etched for 15 seconds with 37%phosphoric acid gel (Condac 37, FGM, Setubal, Portugal). After rinsing for 10 seconds, the etched dentin surfaces were gently dried with an absorbent paper to produce a visibly moist and not desiccated surface. Gluma 2 bond (Heraeus Kulzer, Hanau, Germany) was applied with the applicator tips over the dry dentin surface according to manufacturer’s instructions and light cured for 20 seconds. Resin composite (Charisma diamond, Heraeus-Kulzer, Hanau, Germany) discs were built (Fig. 1d) as two increments by light curing each increment for 20 seconds,with the help of a teflon jig (Ultradent Products Inc., South Jordan, UT) which was adapted perpendicular to the excavated surfaces. The dimensions of the resin composite discs were 2.38 mm in diameter and 2.5 mm in length. The specimens were then transferred to an Instron testing machine (Lloyd, Model LRX, England) and subjected to SBS test. The specimens were positioned in the device so that the shearing stamp would load the composite cylinder at a 90° angle with a crosshead

speed of 1mm/s and a cell load capacity of 1 kN until failure.

Fracture analysis was carried out at X40 magnification under stereomicroscope (Olympus SZ61, Tokyo, Japan). Fractures either in the dental tissue or in the composite was classified as cohesive failure. If dentin tubules were exposed and the residues of either the adhesive and/or the composite were detectable, the specimen was assigned to the group of mixed fractures/failure. Adhesive fracture between adhesive agent and dental hard tissue was classified as adhesive failure.

Statistical analysis: All statistical analyses were carried out using SPSS, version 21.0 (IBM SPSS, Chicago, IL, USA), and the significance level was set to 5% for all tests. Shapiro-Wilk test showed that the microhardness values of the dentin were normally distributed. The comparisons of microhardness and SBS between the FACE and conventionally excavated groups were done using “T test” The frequencies were also calculated for qualitative variables.

**Results**

The mean Vickers microhardness and standard deviations of dentin following conventional excavation and FACE were 70.3±3 and 61.5±5, respectively (Fig. 3, Table 1). No statistically significant difference was found

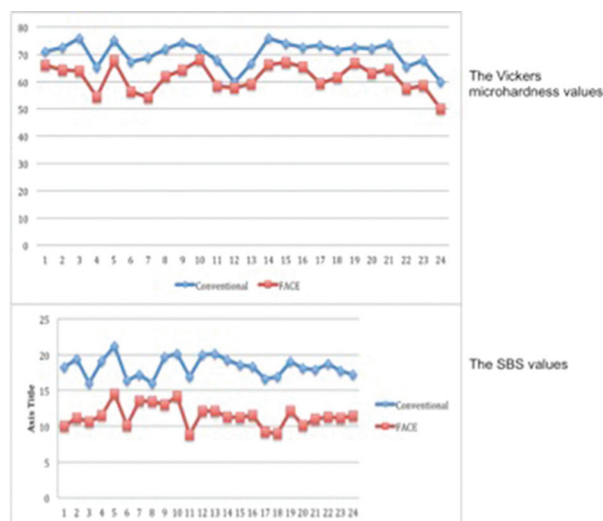


Fig. 3: The Vickers microhardness and SBS values.

Table 1: Mean values of mechanic tests.

	Conventional	FACE
Mechanical Test	X ± SD	X ± SD
Vickers microhardness (VHN)	70.3±3	61.5±5
SBS (MPa)	18.27 ± 1.43	11.42 ± 1.63

between conventional and FACE methods in the microhardness testing ( $p>0.05$ ). The mean SBS of conventional excavation was  $18.27 \pm 1.43$  MPa whereas FACE group exhibited a mean SBS of  $11.42 \pm 1.63$  MPa (Fig. 3, Table 1). Although the conventional excavation yielded to present higher SBS value, the difference between two groups was not statistically significant ( $p>0.05$ ). The results are summarized in Table 1.

The majority of all specimens showed adhesive failure. Only 2 specimens from conventional group showed cohesive failure, which was not significant ( $p>0.05$ ). The results are presented in Table 2.

hardness of dentin following conventional rotary instruments were ranging from 50 to 70 VHN and sound dentin provides 66.94VHN (24-26). In this study, the mean Vickers microhardness of dentin following conventional rotary instrument was found  $70.3 \pm 3$ . Similar to the study of Lai *et al.* (12), present study found that FACE method resulted in significantly lower microhardness than those obtained by conventionally excavated.

In this study, although no significant differences were observed between the microhardness value of two detecting methods, FACE group presented slightly lower microhardness. This decrease may be explained by the se-

**Table 2:** Results of the fracture mode analyses.

Failure mode	FACE		Conventional		p
	n	%	n	%	
Adhesive	24	100	22	92.0	>0.05
Cohesive	-	-	2	8.0	
Mixed	-	-	-	-	
Total	24	100	24	100	

**Discussion**

This study introduced a novel caries-detection method that is potentially capable of objectively differentiating the caries-infected, affected and sound dentin. This method may lead to more precise and minimal sound dentin removal after excavation. Moreover, present study evaluated the excavated dentin areas by the help of dentin microhardness and SBS.

The knowledge of the mechanical properties of human dentin can be helpful to assess the benefits of different treatment alternatives. Microhardness tests, based on the induced permanent surface deformation that remains after removal of the load (17,18), has been used in *in vitro* studies to differentiate the sound and carious dentin or the effects of various types of treatments on dental hard tissues. It offers comparable and objective relative dentine hardness values, which gives direct clinical translational value to operators (19-21). The most commonly used approaches were the Knoop and Vickers hardness tests. Although the optical evaluation of Knoop and Vickers indenter impressions are subjective when transparent materials used such as tooth hard tissues, at lower loads, the data variation errors increase at small measurement and viscoelastic materials exhibit a time dependent elastic recovery leading to additional variations (22). To eliminate these disadvantages, nanoindenters was used to evaluate the nanomechanical properties of dental hard tissues however its high cost limits the its availability (23).

It was previously reported that the mean Vickers micro-

hardness of dentin following conventional rotary instruments were ranging from 50 to 70 VHN and sound dentin provides 66.94VHN (24-26). In this study, the mean Vickers microhardness of dentin following conventional rotary instrument was found  $70.3 \pm 3$ . Similar to the study of Lai *et al.* (12), present study found that FACE method resulted in significantly lower microhardness than those obtained by conventionally excavated. In this study, although no significant differences were observed between the microhardness value of two detecting methods, FACE group presented slightly lower microhardness. This decrease may be explained by the selective removal of caries-infected dentin, leaving more caries-affected dentin tissue, has a lower hardness value. The hardness values at these locations close to the pulp was also reported to be lower compared to superficial dentin, mid-dentin (12,27,28). The selective removal of visualized porphyrins may also lead to the preparation of deeper areas in dentin according to the density and orientation of the dentin tubules where the dentin hardness varies because of different mineralizations. However, variations in the dentin composition and the density of dentin tubules makes bonding to dentin a challenging procedure.

Kumari *et al.* (29) investigated the bond strength of nano-composite resin to superficial/deep dentin and reported a significant decrease in bond strength values from superficial to deep dentin.

In this study, FACE group showed lower SBS results than conventional group, which correlates with the current literature. We also have to take into consideration that, locally more tissue removal via FACE as only bacteria products-contaminated dentin was illuminated and removed and concluded by microhardness test, in this study. The composition of dentin substrate consists of 50% minerals, 20% water, and 30% organic matrix (29). However, it is well known that, as the dentin deepens, this composition changes, number of tubules and amount of water increases. The adhesive resin used in this study contains ethanol as a solvent and after the evaporation of the solvent no movement of liquid must be detected. In the presence of undesirable water arising

in dentin tubules may impair adhesion between composite resin and dentin. Therefore, lower bond strength was achieved in comparison with the superficial dentin. However, although, no difference was observed between the groups in this study, almost all the test specimens showed adhesive fracture in correlation with the current literature (29,30).

Despite the paradigm of minimal-invasive dentistry, the degree of carious dentin that has to be removed from a cavity is still a matter of discussion and it is still an unsolved problem for the practitioner. Yet, it is not known as to how far the dentin must be removed. Although, FACE seems to be promising on minimizing the unnecessary risk of over excavation by detecting only infected dentin and preventing affected dentin, this technique needs to be validated by further *in-vivo/vitro* investigations.

## References

- Fusayama T. Two layers of carious dentin; diagnosis and treatment. *Oper Dent.* 1979;4:63-70.
- McComb D. Systematic review of conservative operative caries management strategies. *J Dent Educ.* 2001;65:1154-61.
- Banerjee A, Watson TF, Kidd EA. Dentine caries excavation: a review of current clinical techniques. *Br Dent J.* 2000;188:476-82.
- Nadanovsky P, Cohen Carneiro F, Souza de Mello F. Removal of caries using only hand instruments: a comparison of mechanical and chemo-mechanical methods. *Caries Res.* 2001;35:384-9.
- Zhang X, Tu R, Yin W, Zhou X, Li X, Hu D. Micro-computerized tomography assessment of fluorescence aided caries excavation (FACE) technology: comparison with three other caries removal techniques. *Aust Dent J.* 2013;58:461-7.
- Kidd EA. How 'clean' must a cavity be before restoration? *Caries Res.* 2004;38:305-13.
- Kidd EA, Joyston-Bechal S, Beighton D. Microbiological validation of assessments of caries activity during cavity preparation. *Caries Res.* 1993;27:402-8.
- Banerjee A, Watson TF, Kidd EA. Dentine caries: take it or leave it? *Dent Update.* 2000;27:272-6.
- Fusayama T, Terachima S. Differentiation of two layers of carious dentin by staining. *J Dent Res.* 1972;51:866.
- Boston DW, Graver HT. Histological study of an acid red caries-disclosing dye. *Oper Dent.* 1989;14:186-92.
- Peskersoy C, Turkun M, Onal B. Comparative clinical evaluation of the efficacy of a new method for caries diagnosis and excavation. *J Conserv Dent.* 2015;18:364-8.
- Lai G, Zhu L, Xu X, Kunzelmann KH. An *in vitro* comparison of fluorescence-aided caries excavation and conventional excavation by microhardness testing. *Clin Oral Investig.* 2014;18:599-605.
- Alfano RR, Yao, S.S. Human teeth with and without dental caries studied by visible luminescent spectroscopy. *J Dent Res.* 1981;60:120-2.
- Lennon AM, Attin T, Martens S, Buchalla W. Fluorescence-aided caries excavation (FACE), caries detector, and conventional caries excavation in primary teeth. *Pediatr Dent.* 2009;31:316-9.
- Lennon AM, Buchalla W, Rassner B, Becker K, Attin T. Efficiency of 4 caries excavation methods compared. *Oper Dent.* 2006;31:551-5.
- Lennon AM, Buchalla W, Switalski L, Stookey GK. Residual caries detection using visible fluorescence. *Caries Res.* 2002;36:315-9.
- Fuentes V, Toledano M, Osorio R, Carvalho RM. Microhardness of superficial and deep sound human dentin. *J Biomed Mater Res A.* 2003;66:850-3.
- Van Meerbeek B, Willems G, Celis JP, Roos JR, Braem M, Lambrechts P, et al. Assessment by nano-indentation of the hardness and elasticity of the resin-dentin bonding area. *J Dent Res.* 1993;72:1434-42.
- Banerjee A, Cook R, Kellow S, Shah K, Festy F, Sherriff M, et al. A confocal micro-endoscopic investigation of the relationship between the microhardness of carious dentine and its autofluorescence. *Eur J Oral Sci.* 2010;118:75-9.
- Almahdy A, Downey FC, Sauro S, Cook RJ, Sherriff M, Richards D, et al. Microbiochemical analysis of carious dentine using Raman and fluorescence spectroscopy. *Caries Res.* 2012;46:432-40.
- Kirsten GA, Takahashi MK, Rached RN, Giannini M, Souza EM. Microhardness of dentin underneath fluoride-releasing adhesive systems subjected to cariogenic challenge and fluoride therapy. *J Dent.* 2010;38:460-8.
- Shahdad SA, McCabe JF, Bull S, Rusby S, Wassell RW. Hardness measured with traditional Vickers and Martens hardness methods. *Dent Mater.* 2007;23:1079-85.
- Marshall GW, Habelitz S, Gallagher R, Balooch M, Balooch G, Marshall SJ. Nanomechanical properties of hydrated carious human dentin. *J Dent Res.* 2001;80:1768-71.
- Fuentes V, Toledano, M., Osorio, R., Carvalho, R.M. Microhardness of superficial and deep sound human dentin. *J Biomed Mater Res.* 2003;66:850-3.
- Garcia-Contreras R, Scougall-Vilchis RJ, Contreras-Bulnes R, Sakagami H, Morales-Luckie RA, Nakajima H. A comparative *in vitro* efficacy of conventional rotatory and chemomechanical caries removal: Influence on cariogenic flora, microhardness, and residual composition. *J Conserv Dent.* 2014;17:536-40.
- Meredith N, Sherriff M, Setchell DJ, Swanson SA. Measurement of the microhardness and Young's modulus of human enamel and dentine using an indentation technique. *Arch Oral Biol.* 1996;41:539-45.
- Craig RG, Peyton, F.A. The microhardness of enamel and dentin. *J Dent Res.* 1958;37:661-8.
- Fusayama T, Okuse K, Hosoda H. Relationship between hardness, discoloration, and microbial invasion in carious dentin. *J Dent Res.* 1966;45:1033-46.
- Kumari RV, Siddaraju K, Nagaraj H, Poluri RK. Evaluation of Shear Bond Strength of Newer Bonding Systems on Superficial and Deep Dentin. *J Int Oral Health.* 2015;7:31-5.
- Pegado RE, do Amaral FL, Florio FM, Basting RT. Effect of different bonding strategies on adhesion to deep and superficial permanent dentin. *Eur J Dent.* 2010;4:110-7.

## Conflict of interest

The authors have declared that no conflict of interest exist.