



ELSEVIER

Available online at www.sciencedirect.com

ScienceDirect

journal homepage: www.e-jds.com



REVIEW ARTICLE

Recent considerations in regenerative endodontic treatment approaches

Hacer Aksel*, Ahmet Serper

Faculty of Dentistry, Hacettepe University, Sıhhiye, Ankara, Turkey

Received 16 October 2013; Final revision received 23 November 2013

Available online 3 June 2014

KEYWORDS

immature permanent teeth;
pulp regeneration;
pulp revascularization;
pulp revitalization;
regenerative endodontics;
tissue engineering

Abstract *Background/purpose:* Regenerative approaches in endodontics are considered in two concepts. One is a revascularization approach to achieve continued apical formation while the other involves the pulp/dentin regeneration via tissue engineering technology. Recently, some case reports have shown that infected immature teeth can be treated by revascularization approach. However, there is still no standardized treatment protocol for this procedure. The purpose of this review article was to evaluate the effects of regenerative endodontic treatment for necrotic immature permanent teeth and to discuss recent treatment approaches.

Materials and methods: Articles published in dental journals from January 2001 to August 2013 were searched using the following keywords: immature permanent teeth OR immature teeth OR pulp revascularization OR pulp revitalization OR regenerative endodontics by using electronic databases (MEDLINE using the PubMed search engine, Embase, Scopus, and Cochrane Central Register of Controlled Trials).

Results: The regenerative endodontic treatments with various methods and materials result in a significant increase in root length and dentinal wall thickness. Stimulation of stem cells in apical root canal system is required to induce tissue formation and continued root development. Alternative disinfection materials and protocols are required.

Conclusion: Although the regenerative treatment approaches have good clinical outcomes in the majority of case reports, the outcomes are unpredictable. Since the current clinical protocols for regenerative endodontics do not fully fulfill the triad of tissue engineering ((growth factors, scaffold and stem cells), further translational studies are required to achieve more pulp- and dentin-like tissue in the root canal system to achieve pulp regeneration.

Copyright © 2014, Association for Dental Sciences of the Republic of China. Published by Elsevier Taiwan LLC. All rights reserved.

* Corresponding author. Department of Endodontics, Faculty of Dentistry, Hacettepe University, 06100 Ankara, Turkey.
E-mail address: hacer.yilmaz@hacettepe.edu.tr (H. Aksel).

Introduction

The main goal of root canal treatment is the prevention or treatment of apical periodontitis.¹ For the treatment of immature permanent teeth, the goal is to restore the original physiologic structures and functions of the pulp–dentin complex. However, treating necrotic immature teeth has always been a clinical challenge for several reasons.² It is difficult to achieve an appropriate apical seal with an open apex by using conventional root canal treatment. In addition, the discontinued development of dentinal walls after pulp necrosis can cause thin dentinal walls that make the tooth more prone to fracture.

One-visit apexification that is performed by placing an apical barrier by using mineral trioxide aggregate (MTA) is an alternative to conventional long-term calcium hydroxide $[\text{Ca}(\text{OH})_2]$ therapy and may shorten the treatment time between the patient's first appointment and the final restoration.³ The survival rate of MTA apexification is greater than that of $\text{Ca}(\text{OH})_2$ apexification.⁴ Mineral trioxide aggregate is effective in supporting the formation of new hard tissue in the apical area of the affected immature necrotic teeth; however, the risk of future fracture may remain because the root width will not increase in MTA apexification-treated teeth.⁴

Regenerative approaches in endodontics comprise two clinical concepts. One concept involves a revitalization approach to achieve tissue regeneration. In this method, new living tissue is expected to form in the cleaned canal space, thereby allowing continued root length and thickness. The other concept is the active pursuit of pulp and dentin regeneration via tissue engineering technology to implant or re-grow pulp tissue. The technology is in its infancy, but it potentially allows immature pulpless teeth to continue growing and maturing. With this understanding, it may be that apexification will become less needed in years to come.

Revascularization is a valuable treatment in immature necrotic teeth. Procedures attempting to preserve the remaining dental pulp stem cells and the mesenchymal stem cells of the apical papilla (SCAPs) can result in root canal revascularization and the completion of root maturation.⁵ Stem cells generally remain in a quiescent state to protect their proliferative potentials *in vivo*.⁶ Quiescent stem cells may be activated by microenvironmental changes such as tissue injury or disease.⁷ In the presence of apical periodontitis, the root canal lumen is probably devoid of vital tissues. However, traces of pulpal tissue may survive apically, even in the presence of a large periapical lesion.^{8,9}

The key procedures of the regenerative protocol are minimal or no instrumentation of the canal while relying on a gentle but thorough irrigation of the root canal system. The disinfection is augmented with intracanal medication, and the treated tooth is sealed with MTA and glass ionomer/resin cement at the completion of the treatment. Periodical follow-ups will take place to observe any continued maturation of the root.¹⁰

To date, pulp revascularization is reported as a promising approach for treating immature permanent teeth. By contrast, there are some drawbacks and variables in

relation to this treatment approach.^{11,12} The purpose of this article was to review the recent literature and to evaluate recent treatment approaches to guide clinicians in using regenerative endodontic procedures in clinical endodontics.

Materials and methods

Articles concerning pulp revascularization published in dental journals from January 2001 to October 2013 were searched using the following keywords: "immature permanent teeth", "immature teeth", "pulp revascularization", "pulp revitalization", or "regenerative endodontics". The electronic databases that use MEDLINE were the PubMed search engine (<http://www.ncbi.nlm.nih.gov/sites/pubmed>), Embase (<http://www.embase.com>), Scopus (<http://www.scopus.com>), and Cochrane Central Register of Controlled Trials (<http://www.cochrane.org>). The data obtained from the clinical case series and case reports are summarized in Table 1.^{4,9,11,12,19,20,23–27,30,31,49} The percentages of the treatment factors are evaluated in Table 2.

Results

Dental caries (12.9%), trauma (33.8%), and *dens evaginatus* (25.9%) were the potential causes of necrotic pulp of immature teeth, that lead to the cessation of root formation. Premolars were the most affected and treated teeth (59.6%). Sodium hypochlorite (2.5–6%) has been used for irrigation and disinfection during regenerative endodontic therapy. Calcium hydroxide and triple antibiotic paste are mostly used as intracanal medicaments. Regenerative endodontic procedures potentially allow the thickening of the dentinal walls (76%) and lengthening of the root canals (54%).

Discussion

Factors causing pulp necrosis in immature teeth

Dental caries, trauma, and anomalous tooth morphology (i.e., *dens evaginatus*) cause pulp necrosis of immature teeth and thereby cause the cessation of root formation. Eradication of bacteria from the pulp canal has a key role in successful revascularization because revascularization halts in the presence of infection.¹³

Premolars are the most affected and treated teeth. This may be related to the prevalence of *dens evaginatus* in this tooth type. *Dens evaginatus* is an uncommon dental anomaly that presents by protrusion of a tubercle from the occlusal surfaces of the posterior teeth or lingual surfaces of the anterior teeth. It occurs primarily in Asian people.¹⁴ It is also called central cusp in premolars.¹⁴ The greatest disadvantage of *dens evaginatus* is that it makes the tooth more susceptible to pulp exposure caused by wear or fracture, and therefore leads to pulpal complications soon after eruption. In addition, because dental trauma mostly occurs in the anterior dental region and the upper incisors are more protracted than the other teeth,¹⁵ these tooth types most often have necrotic dental pulp.

Table 1 The case series and case reports summarized in this review.

Authors	Year	Sex	Tooth type (no.)	Cause (no. of cases)	Change in length	Change in thickness	Follow-up duration (mo)
Jung et al ⁴⁹	2008	5 Females 3 Males	PM (9)	Fracture of the occlusal tubercle (2) Previously initiated therapy (4)	+ (5 cases)	+	24
Shin et al ¹⁹	2009	1 Female	PM	Caries	—	+	19
Ding et al ⁹	2009	2 Females 1 Male	I (2) PM (1)	Caries (2) Dens invaginatus (1)	+ (1 case)	+	17
Chueh et al ²⁵	2009	12 Females 11 Males	I (1) PM (21) M (1)	Caries (1) Trauma (1) Central cusp fracture (21)	+	+	12
Petrino et al ¹²	2010	1 Female 2 Males	I (4) PM (2)	Caries (1) Trauma (4)	+ (4 cases)	+ (4 cases)	10
Kim et al ³⁰	2010	1 Female	I (2)	Trauma (2)	+	+	8
Nosrat et al ²³	2011	1 Female 1 Male	M (2)	Caries	+	+	
Cehreli et al ²⁶	2011	5 Females 1 Male	M (6)	Previously initiated therapy	+	+	9.5
Kim et al ³¹	2012	2 Males	PM (3)		+ (1 case)	+	38
Jeeruphan et al ⁴	2012	10 Females 10 Males	I (7) PM (13)	Caries (1) Dens invaginatus (12) Trauma (7)	+ (6 cases)	+ (3 cases)	21.2
Chen et al ²⁴	2012	11 Females 9 Males	I (10) PM (10)	Caries (3) Dens invaginatus (7) Trauma (10)	+ (15 cases)	+	11.5
Nosrat et al ¹¹	2012	1 Female	I (2)	Trauma	+	—	72
Soares et al ²⁷	2013	1 Female	I	Trauma	+	+	24
Chen et al ²⁰	2013	1 Female	PM	Dens evaginatus	+	+	12

I = incisor; M = molar; PM = premolar.

Disinfection of the root canal system

Decalcification of the dentin surface, removal of the smear layer, exposure of dentinal tubules and collagen fibrils, and release of growth factors from the dentin matrix are required for cellular differentiation at the dentin interface.¹⁶ Sodium hypochlorite (2.5–6%) has been used for irrigation and disinfection during regenerative endodontic therapy. Modification to this protocol—a final step or irrigation with EDTA—has been recommended to optimize the conditions for cellular differentiation, tissue formation, and regeneration.^{17,18} The additional use of chlorhexidine (CHX) has been described in some case reports,^{12,19,20} although it can be detrimental to stem cells.²¹ In addition, the use of the EndoVac system (Discus Dental, Culver City, CA) reportedly provides similar bacterial reduction as that of apical positive pressure irrigation (i.e., conventional irrigation) plus intracanal dressing with a triantibiotic paste, and it has been concluded that using intracanal antibiotics may be unnecessary.²²

In addition to irrigation, the use of intracanal medicaments contributes to the decontamination of root canals. Triple antibiotic paste (TAP),^{11,20,23} calcium hydroxide [Ca(OH)₂],^{24–26} and CHX gel²⁷ have been used for 1–4 weeks for this purpose. For teeth with a persistent infection or in which the canal can not be dried, the medicament

dressing can be repeated until no symptoms or exudation is present.⁹ Redressing the tooth with medicaments may control the infection and eliminate symptoms in time, but this requires stringent follow up. Therefore, noncompliance of the patient may be a contraindication for these procedures. Triple antibiotic pastes are mainly composed of metronidazole, ciprofloxacin, and minocycline. These pastes show antimicrobial activity against endodontic pathogens and satisfactory results with root development in pulp revascularization.²⁸ The pastes nevertheless cause some side effects such as coronal discoloration, bacterial resistance, and allergic reaction.^{29,30} Sealing the dentinal walls of the access cavity by using a dentin bonding agent and a composite resin prior to placing TAP inside the canal has been recommended.²⁹ By contrast, Kim et al³⁰ showed that using dentin bonding agents prior to the placement of TAP does not completely prevent tooth discoloration. Cefaclor is instead recommended to prevent tooth discoloration.³¹

Calcium hydroxide is reportedly a promising alternative for intracanal medications because of its antimicrobial properties,³² the unlikelihood of crown discoloration,²⁵ the possible release of growth factors and biomolecules from dentin,¹⁶ and the availability of this medication in routine clinical practice. Better results are achieved when the placement of Ca(OH)₂ is restricted to the coronal half of the

Table 2 Summary data of the studies included in this review.

Variable	%
Sex	
Female	56.5
Male	43.5
Tooth type	
Incisor	31.3
Premolar	59.6
Molar	9.1
Tooth location	
Maxilla	31.3
Mandible	68.7
Cause	
Caries	12.9
Dens evaginatus	25.9
Trauma	33.8
Fracture	27.4
Intracanal medicament	
Triple antibiotic paste	47
Calcium hydroxide	51
Chlorhexidine gel	2
Change in root length	54
Change in dentin thickness	76
Age (y), mean \pm SD	10.17 \pm 1.96
Follow-up time (mo), mean \pm SD	17.45 \pm 8.58

tooth (53.8% versus 3.3% increase in dentinal wall thickness).³³ However, because of its high pH, Ca(OH)₂ can cause tissue necrosis immediately on contact and thereby destroy tissue differentiation into new pulp.¹⁰ Another disadvantage is associated with the increased risk of root fracture in immature teeth with Ca(OH)₂ dressing because of its reaction with dentin.² A recent study by Ruparel et al³⁴ evaluated the effects of TAP, double antibiotic paste, and Ca(OH)₂ on human SCAPs. Ruparel showed that medicaments used in regenerative procedures have a detrimental effect on the survival of SCAPs, except for Ca(OH)₂. For this reason, it is important to use these medicaments at an adequate concentration to have antibacterial efficacy while not evoking toxicity in the host stem cells. Another recent study reports that mechanical instrumentation of the cervical third and middle third of the root canal with an intracanal dressing composed of Ca(OH)₂ and 2% CHX gel leads to satisfactory root development in necrotic immature teeth.²⁷ Nosrat et al³⁵ recommend using amoxicillin plus clavulanate (i.e., Augmentin, GlaxoSmithKline, Research Triangle Park, NC) as an intracanal medicament for healing periradicular disease. Furthermore, Shin et al¹⁹ present a successful single-visit regenerative endodontic treatment technique to discard the intracanal medication process.

Blood clot formation

Research shows that the inclusion of a blood clot in the root canal tends to improve the revascularization outcome,³⁶ and that the induction of bleeding into the canal may provide stem cells that can induce dentin formation.³⁷

Besides acting as a scaffold, the blood clot may also contain growth and differentiation factors that may be important for successful revascularization of the empty pulp canal.³⁸ Ding et al⁹ suggested that failed regenerative procedures were attributed to the inability to evoke bleeding into the root canal. It is important to consider that if bleeding after canal disinfection is not achieved, clinicians should consider using an anesthetic without a vasoconstrictor when trying to induce bleeding because bleeding is easier when an anesthetic solution does not contain a vasoconstrictor.¹² The bleeding should be allowed to reach a level of 3 mm below the cementoenamel junction, and the tooth is left for 15 minutes so that a blood clot forms.⁹ However, cases of successful revascularization using Ca(OH)₂ without inducing bleeding have also been reported.²⁵ This could be related to the presence of SCAPs in immature teeth, although induced bleeding will increase the chance of stem/progenitor cell migration.

MTA placement

Mineral trioxide aggregate is carefully placed over the blood clot, followed by a wet cotton pellet. A coronal seal with MTA is used because the material possesses an excellent sealing ability.^{39,40} To allow more root development, the coronal edge of the MTA should be placed 1–2 mm apical to the cementoenamel junction rather than 3–4 mm as described by Banchs and Trope.¹⁰ A resorbable barrier is also used to serve as a matrix for the MTA.¹²

The size of the apical foramen

The size of the apical foramen is of great interest, especially for the regenerative endodontic treatment of permanent teeth after the completion of root development. In a radiographic study, Andreasen et al⁴¹ showed that revascularization of the pulp tissue is unpredictable if the apical foramen of the tooth is smaller than 1 mm. However, in a recent animal study, Laureys et al⁴² reported that an apical foramen of 0.32 mm did not prevent the ingrowth of new tissue in two-thirds of the pulp chamber at 90 days after transplantation. With this finding, they concluded that the size of the apical foramen seems not to be the only decisive factor for successful revascularization and ingrowth of new tissue after transplantation. In addition, similar results were described in a case report in which the apical foramen was enlarged only up to 0.6 mm.⁴³

Regenerated tissue properties and tooth vitality

The nature and histologic appearance of the new tissue occurring in the root canal space are also unknown, although a positive response to the pulp sensibility test has been achieved in some patients.⁹ In fact, the lack of response may not even be related to the presence or absence of regenerated nerve tissue. According to Torabinejad and Turman,⁴⁴ the coronal level of regenerated tissue and the thickness of filling materials placed over this tissue both affect the presence or absence of responses to the electric pulp test and cold.

According to a series of clinical and histologic studies, regenerative tissues in root canals are primarily of the following four types: (1) revascularization of the pulp with accelerated dentin formation, which leads to pulp canal obliteration; (2) ingrowth of the cementum and periodontal ligament; (3) ingrowth of the cementum, periodontal ligament, and bone; and (4) ingrowth of bone and bone marrow.⁴⁵ The first type is believed to have the best prognosis. Human teeth with an immature apex are an effective source of cells for hard tissue regeneration.⁴⁶ Hard tissues newly formed on dentinal walls are reportedly distinct from dentin, bone, or bone-like tissue in the root canal space; they resemble cementum, but with significantly different organization and maturation of the collagen matrix.⁴⁷ However, whether the root thickened by cementum-like tissue provides needed physical strength is unknown. Besides the hard tissues, the type of soft tissues newly formed in the root canal is also of interest. In a clinical study,⁴⁸ one patient received a regenerative endodontic treatment by using platelet-rich plasma. Because of the patient's complaint, the root canal treatment was performed. Histologic examination of the tissue removed from the root canal during root canal treatment revealed that it is a pulp-like connective tissue without odontoblasts.⁴⁸

To date, clinical cases have been reported showing that immature teeth healing via revitalization have the characteristics of gaining root thickness and length that resembles the normal maturation of the root.^{4,8,10,19,20,23,24,26,27,29,43,44,49–51} This leads to the speculation that some surviving pulp tissue, and likely the apical papilla, must have been present after disinfection.⁵² Hertwig's epithelial root sheath and the apical papilla have been observed in an immature permanent tooth clinically diagnosed as having irreversible pulpitis.⁵² These vital tissues contribute to the maturation of root development.^{53,54} Inflammation is believed to provide factors that guide the differentiation of stem/progenitor cells in the healing soft tissue into cementoblasts.⁵⁴ However, infection and/or inflammation can hinder the potential of tissue regeneration and stem cell function.⁵⁵

Based on the studies and case reports, Table 3^{8,9,12,17,21,22,30} summarizes some important findings that can guide clinicians to achieve more successful treatment for infected immature permanent teeth. The success rate of regenerative endodontic treatments can be evaluated in accordance with the clinical outcomes. A retrospective outcome study by Jeeruphan et al⁴ showed a great survival rate (100%) with healed cases (80%) and healing cases (20%) after pulp revascularization. This survival rate was based on an approximately 21-month mean follow-up evaluation. Successful outcomes have been reported by clinical cases, histologic analysis revealed tissue repair, instead of regeneration, of the pulp-dentin complex.⁵⁴ In a recent case report, Nosrat et al³⁵ showed the maturation of the root apex in the absence of the regenerated pulp tissue in the root canal by observing the root canal with an operation microscope after patients had undergone 17 months of initial endodontic therapy. Therefore, it could be concluded that the protocol for regenerative endodontic treatment is not predictable for pulp-dentin regeneration. To achieve the regeneration of anatomic pulp-dentin

Table 3 Recommendations for successful regenerative endodontic treatment, based on the reviewed articles.

Authors	Year	Important findings
Ding et al ⁹	2009	Case selection is the most important part of this treatment approach because it requires stringent follow-up.
Cotti et al ⁸	2008	Concentrations of NaOCl ranging from 2.5% to 6% have been used for irrigation and provides favorable results.
Kim et al ³⁰	2010	
Galler et al ¹⁷	2011	A final irrigation with EDTA provides more optimal conditions for cellular differentiation, tissue formation, and regeneration.
Trevino et al ²¹	2011	CHX is detrimental to stem cells of the apical papilla. It is not recommended.
Cohenca et al ²²	2010	EndoVac system (Discus Dental, Culver City, CA) can be used without the necessity of intracanal medication (especially triantibiotic paste) to prevent further discoloration.
Ding et al ⁹	2009	If Ca(OH) ₂ is used as an intracanal medicament, its placement should be restricted to the coronal half of the tooth to achieve greater increase in the dentinal wall thickness.
Petrino et al ¹²	2010	If bleeding after root canal disinfection fails, clinicians should consider using an anesthetic without a vasoconstrictor when trying to induce bleeding.
Ding et al ⁹	2009	For the formation of a blood clot, the tooth should be left for 15 minutes. The blood clot acts as a suitable barrier for the placement of MTA (ProRoot MTA, Dentsply Maillefer, Tulsa, OK, USA).
Petrino et al ¹²	2010	It is recommended to place the coronal edge of the MTA (ProRoot MTA, Dentsply Maillefer, Tulsa, OK, USA) 1–2 mm apical to the CEJ to allow new tissue growth into the root canal.

Ca(OH)₂ = calcium hydroxide; CEJ = cementoenamel junction; CHX = chlorhexidine; MTA = mineral triaggregate; NaOCl = sodium hypochlorite.

complex, it is necessary to add growth factors and scaffolds. Utilizing of growth factors to regenerate pulp tissue without the inclusion of cells yielded the formation of regenerated dental-pulp-like tissue in endodontically treated root canals.⁵⁶ This approach, called "cell homing", contains active recruitment of endogenous cells such as stem/progenitor cells into an anatomic compartment.⁵⁷ The regeneration of dental pulp by cell homing, rather than by cell delivery, can accelerate clinical translation, although stem cell-mediated tissue regeneration has more optimal results and has the capability of *de novo* regeneration of pulp and new dentin.^{58,59} The success of applying cell homing and cell-based tissue engineering to regenerate pulp and dentin in the root canal system is based on pre-clinical studies. For this reason, more translational research is required to evaluate the effects of growth factors, stem cells, and scaffolds in treating immature permanent tooth with pulp necrosis.

Conclusion

Clinical literature indicates that regenerative endodontic treatments using various methods and materials result in a significant increase in root length and dentinal wall thickness. This clinical approach is technically simple and can be completed without expensive biotechnology by using currently available instruments and medicaments. However, for regenerative endodontic procedures to be widely available and predictable, it seems essential to focus on tissue engineering therapies to regenerate pulp-dentin tissue. The proposed therapy can provide a tissue-engineered pulp construct to implant into the root canal. However, this approach is still in its infancy and more translational research is needed to continue to improve this method.

Conflicts of interest

The authors have no conflicts of interest relevant to this article.

References

- Ørstavik D, Pitt Ford T. Apical periodontitis: microbial infection and host responses. In: *Essential Endodontontology: Prevention and Treatment of Apical Periodontitis*. Oxford: Blackwell Science, 1998:1–8.
- Andreasen JO, Farik B, Munksgaard EC. Long-term calcium hydroxide as a root canal dressing may increase risk of root fracture. *Dent Traumatol* 2002;18:134–7.
- Hatibovic-Kofman S, Raimundo L, Zheng L, Chong L, Friedman M, Andreasen JO. Fracture resistance and histological findings of immature teeth treated with mineral trioxide aggregate. *Dent Traumatol* 2008;24:272–6.
- Jeeruphan T, Jantarat J, Yanpiset K, Suwannapan L, Khewsawai P, Hargreaves KM. Mahidol study 1: comparison of radiographic and survival outcomes of immature teeth treated with either regenerative endodontic or apexification methods: a retrospective study. *J Endod* 2012;38:1330–6.
- Sonoyama W, Liu Y, Yamaza T, et al. Characterization of the apical papilla and its residing stem cells from human immature permanent teeth: a pilot study. *J Endod* 2008;34:166–71.
- Sang L, Coller HA, Roberts JM. Control of the reversibility of cellular quiescence by the transcriptional repressor HES1. *Science* 2008;321:1095–100.
- Mitsiadis TA, Barrandon O, Rochat A, Barrandon Y, De Bari C. Stem cell niches in mammals. *Exp Cell Res* 2007;313:3377–85.
- Cotti E, Mereu M, Lusso D. Regenerative treatment of an immature, traumatized tooth with apical periodontitis: report of a case. *J Endod* 2008;34:611–6.
- Ding RY, Cheung GS, Chen J, Yin XZ, Wang QQ, Zhang CF. Pulp revascularization of immature teeth with apical periodontitis: a clinical study. *J Endod* 2009;35:745–9.
- Banchs F, Trope M. Revascularization of immature permanent teeth with apical periodontitis: new treatment protocol? *J Endod* 2004;30:196–200.
- Nosrat A, Homayounfar N, Oloomi K. Drawbacks and unfavorable outcomes of regenerative endodontic treatments of necrotic immature teeth: a literature review and report of a case. *J Endod* 2012;38:1428–34.
- Petrino JA, Boda KK, Shambarger S, Bowles WR, McClanahan SB. Challenges in regenerative endodontics: a case series. *J Endod* 2010;36:536–41.
- Love RM. Bacterial penetration of the root canal of intact incisor teeth after a simulated traumatic injury. *Endod Dent Traumatol* 1996;12:289–93.
- Levitin ME, Himel VT. *Dens evaginatus*: literature review, pathophysiology, and comprehensive treatment regimen. *J Endod* 2006;32:1–9.
- Bastone EB, Freer TJ, McNamara JR. Epidemiology of dental trauma: a review of the literature. *Aust Dent J* 2000;45:2–9.
- Graham L, Cooper PR, Cassidy N, Nor JE, Sloan AJ, Smith AJ. The effect of calcium hydroxide on solubilisation of bio-active dentine matrix components. *Biomaterials* 2006;27:2865–73.
- Galler KM, D'Souza RN, Federlin M, et al. Dentin conditioning codetermines cell fate in regenerative endodontics. *J Endod* 2011;37:1536–41.
- Pang N-S, Lee SJ, Kim E, et al. Effect of EDTA on attachment and differentiation of dental pulp stem cells. *J Endod*. in press, <http://dx.doi.org/10.1016/j.joen.2013.09.007>
- Shin SY, Albert JS, Mortman RE. One step pulp revascularization treatment of an immature permanent tooth with chronic apical abscess: a case report. *Int Endod J* 2009;42:1118–26.
- Chen X, Bao ZF, Liu Y, Liu M, Jin XQ, Xu XB. Regenerative endodontic treatment of an immature permanent tooth at an early stage of root development: a case report. *J Endod* 2013;39:719–22.
- Trevino EG, Patwardhan AN, Henry MA, et al. Effect of irrigants on the survival of human stem cells of the apical papilla in a platelet-rich plasma scaffold in human root tips. *J Endod* 2011;37:1109–15.
- Cohena N, Heilborn C, Johnson JD, Flores DS, Ito IY, da Silva LA. Apical negative pressure irrigation versus conventional irrigation plus triantibiotic intracanal dressing on root canal disinfection in dog teeth. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2010;109:e42–6.
- Nosrat A, Seifi A, Asgary S. Regenerative endodontic treatment (revascularization) for necrotic immature permanent molars: a review and report of two cases with a new biomaterial. *J Endod* 2011;37:562–7.
- Chen MY, Chen KL, Chen CA, Tayebaty F, Rosenberg PA, Lin LM. Responses of immature permanent teeth with infected necrotic pulp tissue and apical periodontitis/abscess to revascularization procedures. *Int Endod J* 2012;45:294–305.
- Chueh LH, Ho YC, Kuo TC, Lai WH, Chen YH, Chiang CP. Regenerative endodontic treatment for necrotic immature permanent teeth. *J Endod* 2009;35:160–4.

26. Cehreli ZC, Isbitiren B, Sara S, Erbas G. Regenerative endodontic treatment (revascularization) of immature necrotic molars medicated with calcium hydroxide: a case series. *J Endod* 2011;37:1327–30.
27. Soares Ade J, Lins FF, Nagata JY, et al. Pulp revascularization after root canal decontamination with calcium hydroxide and 2% chlorhexidine gel. *J Endod* 2013;39:417–20.
28. Windley 3rd W, Teixeira F, Levin L, Sigurdsson A, Trope M. Disinfection of immature teeth with a triple antibiotic paste. *J Endod* 2005;31:439–43.
29. Reynolds K, Johnson JD, Cohenca N. Pulp revascularization of necrotic bilateral bicuspids using a modified novel technique to eliminate potential coronal discolouration: a case report. *Int Endod J* 2009;42:84–92.
30. Kim JH, Kim Y, Shin SJ, Park JW, Jung IY. Tooth discoloration of immature permanent incisor associated with triple antibiotic therapy: a case report. *J Endod* 2010;36:1086–91.
31. Kim DS, Park HJ, Yeom JH, et al. Long-term follow-ups of revascularized immature necrotic teeth: three case reports. *Int J Oral Sci* 2012;4:109–13.
32. Lana PE, Scelza MF, Silva LE, Mattos-Guaraldi AL, Hirata Junior R. Antimicrobial activity of calcium hydroxide pastes on *Enterococcus faecalis* cultivated in root canal systems. *Braz Dent J* 2009;20:32–6.
33. Bose R, Nummikoski P, Hargreaves K. A retrospective evaluation of radiographic outcomes in immature teeth with necrotic root canal systems treated with regenerative endodontic procedures. *J Endod* 2009;35:1343–9.
34. Ruparel NB, Teixeira FB, Ferraz CC, Diogenes A. Direct effect of intracanal medicaments on survival of stem cells of the apical papilla. *J Endod* 2012;38:1372–5.
35. Nosrat A, Li KL, Vir K, Hicks ML, Fouad AF. Is pulp regeneration necessary for root maturation? *J Endod* 2013;39:1291–5.
36. Thibodeau B, Teixeira F, Yamauchi M, Caplan DJ, Trope M. Pulp revascularization of immature dog teeth with apical periodontitis. *J Endod* 2007;33:680–9.
37. Lovelace TW, Henry MA, Hargreaves KM, Diogenes A. Evaluation of the delivery of mesenchymal stem cells into the root canal space of necrotic immature teeth after clinical regenerative endodontic procedure. *J Endod* 2011;37:133–8.
38. Freymiller EG. Platelet-rich plasma: evidence to support its use. *J Oral Maxillofac Surg* 2004;62:1046. author reply 47–8.
39. Fridland M, Rosado R. Mineral trioxide aggregate (MTA) solubility and porosity with different water-to-powder ratios. *J Endod* 2003;29:814–7.
40. Xavier CB, Weismann R, de Oliveira MG, Demarco FF, Pozza DH. Root-end filling materials: apical microleakage and marginal adaptation. *J Endod* 2005;31:539–42.
41. Andreasen JO, Paulsen HU, Yu Z, Bayer T, Schwartz O. A long-term study of 370 autotransplanted premolars. Part II. Tooth survival and pulp healing subsequent to transplantation. *Eur J Orthod* 1990;12:14–24.
42. Laureys WG, Cuvelier CA, Dermaut LR, De Pauw GA. The critical apical diameter to obtain regeneration of the pulp tissue after tooth transplantation, replantation, or regenerative endodontic treatment. *J Endod* 2013;39:759–63.
43. Paryani K, Kim SG. Regenerative endodontic treatment of permanent teeth after completion of root development: a report of 2 cases. *J Endod* 2013;39:929–34.
44. Torabinejad M, Turman M. Revitalization of tooth with necrotic pulp and open apex by using platelet-rich plasma: a case report. *J Endod* 2011;37:265–8.
45. Andreasen JO, Bakland LK. Pulp regeneration after non-infected and infected necrosis, what type of tissue do we want? A review. *Dent Traumatol* 2012;28:13–8.
46. Abe S, Yamaguchi S, Watanabe A, Hamada K, Amagasa T. Hard tissue regeneration capacity of apical pulp derived cells (APDCs) from human tooth with immature apex. *Biochem Biophys Res Commun* 2008;371:90–3.
47. Yamauchi N, Nagaoka H, Yamauchi S, Teixeira FB, Miguez P, Yamauchi M. Immunohistological characterization of newly formed tissues after regenerative procedure in immature dog teeth. *J Endod* 2011;37:1636–41.
48. Torabinejad M, Faras H. A clinical and histological report of a tooth with an open apex treated with regenerative endodontics using platelet-rich plasma. *J Endod* 2012;38:864–8.
49. Jung IY, Lee SJ, Hargreaves KM. Biologically based treatment of immature permanent teeth with pulpal necrosis: a case series. *J Endod* 2008;34:876–87.
50. Ritter AL, Ritter AV, Murrah V, Sigurdsson A, Trope M. Pulp revascularization of replanted immature dog teeth after treatment with minocycline and doxycycline assessed by laser Doppler flowmetry, radiography, and histology. *Dent Traumatol* 2004;20:75–84.
51. Thomson A, Kahler B. Regenerative endodontics—biologically-based treatment for immature permanent teeth: a case report and review of the literature. *Aust Dent J* 2010;55:446–52.
52. Shimizu E, Jong G, Partridge N, Rosenberg PA, Lin LM. Histologic observation of a human immature permanent tooth with irreversible pulpitis after revascularization/regeneration procedure. *J Endod* 2012;38:1293–7.
53. Huang GT, Sonoyama W, Liu Y, Liu H, Wang S, Shi S. The hidden treasure in apical papilla: the potential role in pulp/dentin regeneration and bioroot engineering. *J Endod* 2008;34:645–51.
54. Wang X, Thibodeau B, Trope M, Lin LM, Huang GT. Histologic characterization of regenerated tissues in canal space after the revitalization/revascularization procedure of immature dog teeth with apical periodontitis. *J Endod* 2010;36:56–63.
55. Thomas MV, Puleo DA. Infection, inflammation, and bone regeneration: a paradoxical relationship. *J Dent Res* 2011;90:1052–61.
56. Kim JY, Xin X, Moioli EK, et al. Regeneration of dental-pulp-like tissue by chemotaxis-induced cell homing. *Tissue Eng Part A* 2010;16:3023–31.
57. Mao JJ, Stosich MS, Moioli EK, et al. Facial reconstruction by biosurgery: cell transplantation versus cell homing. *Tissue Eng Part B Rev* 2010;16:257–62.
58. Galler KM, Hartgerink JD, Cavender AC, Schmalz G, D’Souza RN. A customized self-assembling peptide hydrogel for dental pulp tissue engineering. *Tissue Eng Part A* 2012;18:176–84.
59. Huang GT, Yamaza T, Shea LD, et al. Stem/progenitor cell-mediated de novo regeneration of dental pulp with newly deposited continuous layer of dentin in an in vivo model. *Tissue Eng Part A* 2010;16:605–15.