

Epidemiology of penicillin resistance in *Streptococcus pneumoniae* isolates in Kayseri, Turkey

D. Esel¹, B. Sümerkan¹ and S. Kocagöz²

¹Department of Microbiology, University of Erciyes, Kayseri, Turkey and ²Department of Infectious Diseases, University of Hacettepe, Ankara, Turkey

Objective To determine the penicillin resistance and serotype distribution of *Streptococcus pneumoniae* strains and to identify clonal relationships of isolates resistant to penicillin by means of pulsed-field gel electrophoresis (PFGE).

Methods In total, 193 *S. pneumoniae* strains were isolated from clinical specimens between November 1997 and January 2000. Susceptibility testing was carried out by E test, and serotyping by the Quellung reaction. Clonal relationship was analyzed by using PFGE with *smal* endonuclease.

Results Of the *S. pneumoniae* isolates, 23% were intermediately resistant to penicillin. There were no high-level resistant pneumococci. The majority of isolates intermediately resistant to penicillin were of serogroups/serotypes 19, 23, 14 and 1, in descending order of frequency. There were eight major clones in strains intermediately resistant to penicillin. It was seen that serogroups in the 23-valent polysaccharide vaccine, 7-valent, 9-valent, and 11-valent vaccine formulations caused 92%, 75%, 78% and 87% of pneumococcal diseases in our region, respectively.

Conclusion Penicillin resistance in *S. pneumoniae* is relatively uncommon in Kayseri. All vaccine formulations can prevent the majority of pneumococcal diseases, and there is genetic heterogeneity in intermediately penicillin-resistant pneumococci in this region.

Keywords *Streptococcus pneumoniae*, penicillin resistance, serotyping, PFGE

Accepted 3 July 2001

Clin Microbiol Infect 2001; 7: 548–552

INTRODUCTION

Streptococcus pneumoniae is the leading bacterial cause of community-acquired pneumonia as well as acute otitis media, sinusitis, and meningitis. Despite the advent of penicillin and effective antimicrobial agents, and the availability of vaccine, pneumococcal infections are still associated with high morbidity and mortality rates [1,2]. Since the first description of a strain with diminished susceptibility in 1967, resistance among *Streptococcus pneumoniae* isolates has rapidly spread throughout the world [3,4].

Clonal spread and horizontal gene transfer are the plausible explanations for the rapid emergence of penicillin-resistant *Streptococcus pneumoniae* [5]. Clonal spread is the cause of spread of resistant strains between different countries and continents [6,7].

Prevention of pneumococcal disease with vaccine is very important, particularly for individuals in high-risk groups, to decrease morbidity and mortality. The currently available

pneumococcal polysaccharide vaccine is not protective in children under 2 years old and in immunosuppressed patients [8]. To improve the current pneumococcal polysaccharide vaccines, new conjugate vaccines in which polysaccharides are covalently linked to carrier proteins have been developed [9].

It is very important to know the serotype distribution and antibiotic resistance in a particular area in order to estimate the probable efficacy of the vaccines in that region [10,11].

The aims of this study were to determine the serotype distribution and penicillin resistance of *Streptococcus pneumoniae* strains in Kayseri and to identify clonal relationships of penicillin-resistant strains by means of pulsed-field gel electrophoresis (PFGE).

MATERIALS AND METHODS

Bacterial strains

The pneumococcal strains were isolated from 193 patients in the Department of Microbiology of Erciyes University Hospital in Kayseri, a city in central Turkey with a population of about 800 000. All of the isolates were collected from patients with community-acquired infection. The study was performed

Corresponding author and reprint requests: D. Esel, Department of Microbiology, University of Erciyes, Kayseri 38039, Turkey
Tel: +90 (352) 4374937 2472
Fax: +90 (352) 437 52 88 52

over a period of 26 months (November 1997 to January 2000). The strains were isolated from different specimen sources: 49 from sputum, 34 from cerebrospinal fluid, 33 from ear discharge, 26 from blood infections, 22 from ocular samples, 10 from pleural fluid, eight from soft tissue, seven from nasotracheal aspirate, two from bronchoalveolar lavage, and one from peritoneal fluid.

Identification was carried out by colony morphology on blood agar, Gram stain, susceptibility to optochin, and bile solubility [12]. Strains were stored in microbanks at -70°C until analyzed.

Streptococcus pneumoniae ATCC 49619 was used as the quality control strain for susceptibility testing, and *Staphylococcus aureus* NCTC 8325 to ensure the comparability of the gels during PFGE.

Sensitivity to penicillin G

Susceptibility testing was performed by E test (AB Biodisk, Solna, Sweden) on Mueller–Hinton agar with 5% sheep blood and incubation in the presence of 5% CO_2 . The E test was performed according to the manufacturer's instructions. Strains with $\text{MIC} \leq 0.06 \text{ mg/L}$ were considered to be susceptible, those with MICs between 0.1 and 1.0 mg/L intermediate, and those with $\text{MIC} \geq 2.0 \text{ mg/L}$ highly resistant, according to the NCCLS protocol [13].

Serotyping

Serotyping was performed by the Quellung reaction with Pneumotest (Statens Serum Institute, Copenhagen, Denmark).

Pulsed-field gel electrophoresis

This part of the study was performed in Hacettepe University Faculty of Medicine, Department of Infectious Diseases,

Ankara, Turkey. PFGE was carried out on intermediately resistant isolates ($n = 44$). *Streptococcus pneumoniae* DNA embedded in agarose blocks was prepared as described by Soares [6]. For digestion, restriction endonuclease *smI* (Promega, Madison, WI, USA) was used. The DNA macrorestriction fragments were separated in 1.1% agarose gel (Genaxis, Spechbach, Germany) by PFGE (General Navigator, Pharmacia, Uppsala, Sweden). λ -Ladder (Sigma, Deisenhofen, Germany) was used as a molecular weight marker.

Analysis of PFGE profiles

Visual comparison of macrorestriction patterns of the chromosomal DNAs was done using the 'Alpha Imager Documentation and Analysis System' (USA), and distinct patterns were assigned an arbitrary PFGE designation. Assuming that a single mutational event in the chromosomal DNA could introduce maximally a three-fragment difference in the restriction pattern, strains showing more than three-fragment variations were assumed to represent major patterns (assignment of capital letters), while one- to three-fragment differences were considered to represent subtypes (capital letters with numerical subcode) [14].

RESULTS

Intermediate-level penicillin resistance was observed in 44 (23%) of the 193 strains, while there was no strain highly resistant to penicillin. MIC range, MIC_{50} and MIC_{90} (mg/L) of the strains for penicillin were 0.004–1, 0.01, and 0.25, respectively.

The serogroup/serotype distribution of 193 strains is shown in Figure 1. Fifteen strains (8%) could not be serotyped with Pneumotest. It was seen that the majority of isolates were of

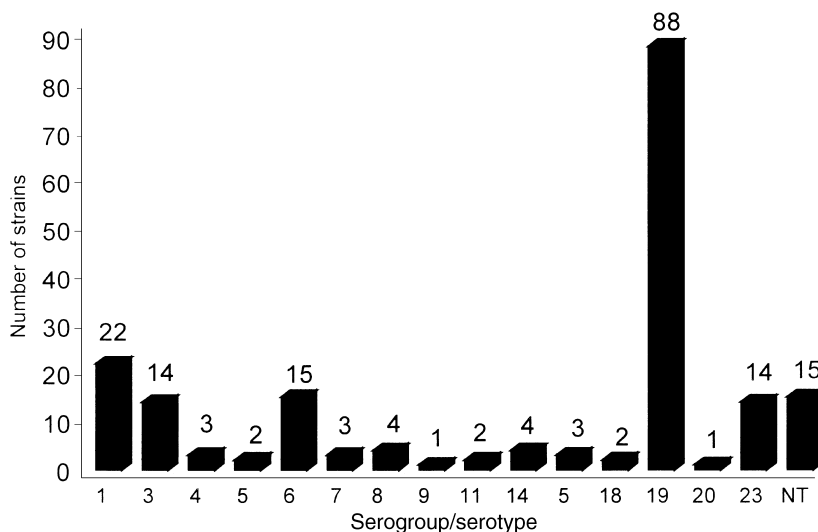


Figure 1 Serogroup/serotype distribution of 193 *Streptococcus pneumoniae* strains.

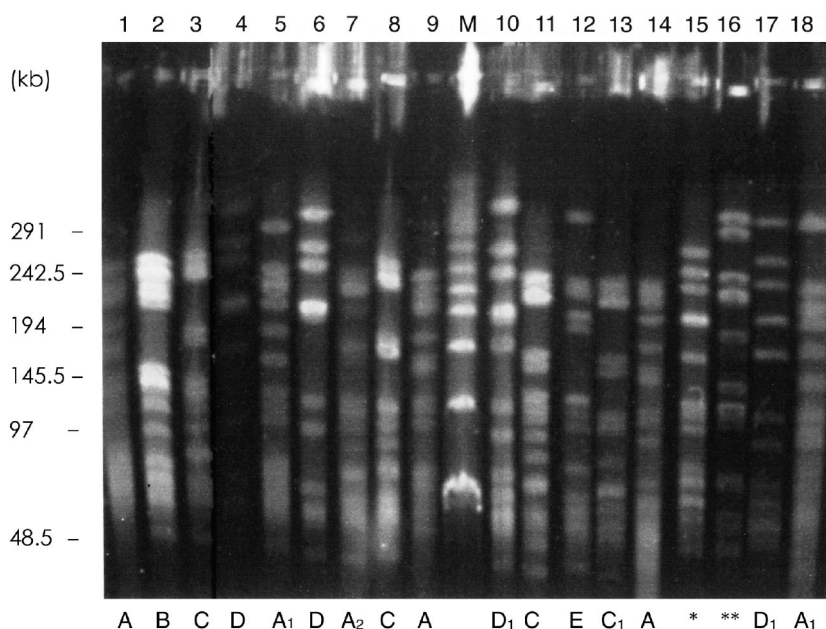


Figure 2 Representative patterns of the predominant PFGE types. M, molecular weight marker (kb). **Staphylococcus aureus* NCTC 8325; ***Streptococcus pneumoniae* ATCC 49619.

serogroups/serotypes 19, 1, 6, 23 and 3, in descending order of frequency. Among the 44 intermediately resistant strains, the most predominant serogroups/serotypes were found to be 19 ($n=22$), 23 ($n=11$), 14 ($n=3$), and 1 ($n=3$).

The clonal relationships of isolates that were intermediately resistant to penicillin were analyzed by means of PFGE. In these strains, eight major patterns (A–H) were observed. The majority of the strains (41%) belonged to PFGE type A; however, there were three different subtypes in clone A. Representative patterns of the predominant PFGE types are shown in Figure 2. No relationship was found between serotypes and PFGE types. Genetic heterogeneity was seen in strains intermediately resistant to penicillin.

DISCUSSION

Shortly after the isolation of pneumococci with moderate penicillin resistance in Australia and New Guinea [15], penicillin-resistant pneumococci emerged in many countries. Today, the prevalence of pneumococci resistant to penicillin is increasing worldwide, particularly in Spain [16,17], Hungary [18], and South Africa [19]. Antimicrobial use is generally considered to be one of the major factors in resistance [20,21]. Although antimicrobial use is relatively unrestricted, and self-administration is common in Kayseri, it is surprising that there is still no highly resistant pneumococcal strain and no increase in the prevalence of intermediate-level resistance from 1992 to 2000 in this region [22,23].

It is known that serotype distribution of strains causing invasive disease, nasopharyngeal colonisation and antibiotic

resistance is related to age, geography, and socio-economic conditions. Analysis of the leading serotypes in a particular area is very important to evaluate the efficacy of vaccines [10,11].

Penicillin resistance is restricted worldwide to a few serogroups. These serogroups are 6, 9, 14, 19, and 23 [24–27]. However, there is a much wider range in serogroups of the strains intermediately resistant to penicillin [28]. In this study, we found that the leading serogroups/serotypes were 19, 1 and 6 in Kayseri. Intermediate-level resistance to penicillin was mostly observed in serogroups 19 and 23. It was observed that serogroups in the 23-valent-polysaccharide vaccine, 7-valent, 9-valent and 11-valent vaccine formulations caused 92%, 75%, 78% and 87% of pneumococcal diseases in Kayseri, respectively. Each formulation can be used and prevent most of the pneumococcal diseases in this region.

There are three hypotheses about the rapid spread of penicillin resistance:

1. The altered penicillin-binding protein (PBP) genes arise by interspecies recombinational events in which segments of native PBP genes are replaced with the corresponding segments from related streptococcal species [29,30].
2. Horizontal transfer of altered PBP genes from resistant strains to susceptible strains is the main cause of spread of resistance [31].
3. The importation and clonal spread of a small number of resistant clones are important factors in the global increase in the incidence of penicillin-resistant pneumococci [6,7].

In general, it can be proposed that resistant pneumococci that are not closely related genetically but that contain identical altered PBP genes have arisen by horizontal spread, whereas

isolates that are genetically indistinguishable are the result of clonal spread [5]. Clonal spread is the most important mechanism, and causes the spread of resistant clones to geographically distinct areas [6,7].

Spanish serotype 23F strains [32] resistant to penicillin, tetracycline, and chloramphenicol, and variably resistant to erythromycin, have been described as a distinct clone that has spread to numerous countries in Europe [6,31,33], the USA [32,34], and South Africa [35]. The same bacterial clone, but with a 19F capsule, has been found in many countries, suggesting capsular switching [31,36,37]. Serotype 23F clones unique to certain areas have been identified in Finland [38], Italy [37], Israel [39], and Bulgaria [40]. Nevertheless, in Japan and Korea, genetic heterogeneity has been reported in serotype 23F strains resistant to penicillin [41,42]. Not only the 23F clone, but also the 14 [33,40], 9V [31,37] and 19A [18,40] clones, have been dispersed throughout the world.

Although resistant strains have highly related PFGE patterns, there is genetic heterogeneity in intermediate and susceptible strains [38,43]. We found eight different major clones, of which 41% belonged to PFGE type A in our strains.

In conclusion, there are still no high-level penicillin-resistant pneumococci in Kayseri. All of the vaccine formulations are sufficient to prevent most of the pneumococcal diseases in this region. There is no relationship between PFGE type and serotype, and there is genetic heterogeneity in pneumococci intermediately resistant to penicillin.

REFERENCES

- Musher DM. *S. pneumoniae*. In: Mandell GL, Douglas RG, Bennett JE, Dolin R, eds. *Principles and Practice of Infectious Diseases*, 4th edn. New York: Churchill Livingstone, 1995: 1811–26.
- Caputo GM, Appelbaum PC, Liu HH. Infections due to penicillin-resistant pneumococci: clinical, epidemiologic and microbiologic features. *Arch Intern Med* 1993; 153: 1301–10.
- Tomasz A. Antibiotic resistance in *Streptococcus pneumoniae*. *Clin Infect Dis* 1997; 24(suppl 1): 85–8.
- Appelbaum PC. Antimicrobial resistance in *Streptococcus pneumoniae*: an overview. *Clin Infect Dis* 1992; 15: 77–83.
- Doit C, Denamur E, Picard B, Geslin P, Elion J, Bingen E. Mechanisms of the spread of penicillin resistance in *Streptococcus pneumoniae* strains causing meningitis in children in France. *J Infect Dis* 1996; 174: 520–8.
- Soares S, Kristinsson KG, Musser JM, Tomasz A. Evidence for the introduction of a multiresistant clone of serotype 6B *Streptococcus pneumoniae* from Spain to Iceland in the late 1980s. *J Infect Dis* 1993; 168: 158–63.
- Muñoz R, Musser JM, Crain M et al. Geographic distribution of penicillin-resistant clones of *Streptococcus pneumoniae*: characterization by penicillin-binding protein profile, surface protein A typing and multilocus enzyme analysis. *Clin Infect Dis* 1992; 15: 112–18.
- Siber GR. Pneumococcal disease: prospects for a new generation of vaccines. *Science* 1994; 265: 1385–7.
- Klein DL, Eskola J. Development and testing of *Streptococcus pneumoniae* conjugate vaccines. *Clin Microbiol Infect* 1999; 5(suppl 4): S17–28.
- Block SI, Harrison JA, Hedrick RD et al. Penicillin-resistant *Streptococcus pneumoniae* in acute otitis media: risk factors, susceptibility patterns and antimicrobial management. *Pediatr Infect Dis J* 1995; 14: 751–9.
- Boken DJ, Chartrand SA, Moland ES, Goering RV. Colonization with penicillin-nonsusceptible *Streptococcus pneumoniae* in urban and rural child-care centers. *Pediatr Infect Dis J* 1996; 15: 667–72.
- Pratt-Rippin K, Pezzlo M. Identification of gram-positive bacteria. In: Isenberg HD, ed. *Clinical Microbiology Procedures Handbook*, Vol. 1. Washington DC: ASM Press, 1992: 1.20.19–1.20.20.
- National Committee for Clinical Laboratory Standards. *Performance Standards for Antimicrobial Susceptibility Testing*; Ninth Informational Supplement. NCCLS document M100-S9. Wayne Pa: NCCLS, 1999.
- Tenover FC, Arbeit RD, Goering RV et al. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J Clin Microbiol* 1995; 33: 2233–9.
- Hansman D, Bullen MM. A resistant pneumococcus. *Lancet* 1967; 2: 264–5.
- Baquero F, García-Rodríguez JH, de García Lomas J et al. Antimicrobial resistance of 1113 *Streptococcus pneumoniae* isolates from patients with respiratory tract infections in Spain: results of a 1-year (1996–1997) multicenter surveillance study. *Antimicrob Agents Chemother* 1999; 43: 357–9.
- Linares J, Tubau F, Domínguez MA. Antibiotic resistance in *Streptococcus pneumoniae* in Spain: an overview of the 1990s. In: Tomasz A, ed. *Streptococcus Pneumoniae: Molecular Biology and Mechanisms of Disease*, 1st edn. Larchmont: Mary Ann Liebert Inc., 2000: 399–407.
- Marton A, Gulyas M, Munoz R, Tomasz A. Extremely high incidence of antibiotic resistance in clinical isolates of *Streptococcus pneumoniae* in Hungary. *J Infect Dis* 1991; 163: 542–8.
- Klugman K. Pneumococcal resistance to antibiotics. *Clin Microbiol Rev* 1990; 3: 171–96.
- Nava JM, Bella F, Garau J et al. Predictive factors for invasive disease due to penicillin-resistant *Streptococcus pneumoniae*: a population based study. *Clin Infect Dis* 1994; 19: 884–90.
- Castillo F, Baquero F, Garcia A. Influence of recent antibiotic therapy on antimicrobial resistance of *Streptococcus pneumoniae* in children with acute otitis media in Spain. *Pediatr Infect Dis J* 1998; 17: 94–7.
- Sümerkan B, Aygen B, Öztürk M, Doganay M. Pnömonok enfeksiyonları ve penisilin direnci. *Klinik Derg* 1993; 6: 29–30.
- Sümerkan B, Gökahmetoğlu S, Aygen B, Kocagöz S. Klinik örneklerden izole edilen *Streptococcus pneumoniae* suslarının çeşitli antibiyotiklere duyarlılıkları. *Mikrobiyol Bült* 1997; 31: 331–8.
- Bédos JP, Chevret S, Chastang C, Geslin P, Régnier B, the French Cooperative Pneumococcus Study Group. Epidemiological features of and risk factors for infection by *Streptococcus pneumoniae* strains with diminished susceptibility to penicillin: findings of a French survey. *Clin Infect Dis* 1996; 22: 63–72.
- Talon D, Mulin B, Dupont MJ et al. Epidemiology of penicillin resistance in *Streptococcus pneumoniae* isolates in eastern France. *Clin Microbiol Infect* 1998; 4: 11–17.
- Scott JAG, Hall AJ, Hannington A et al. Serotype distribution and prevalence of resistance to benzylpenicillin in three representative populations of *Streptococcus pneumoniae* isolates from the coast of Kenya. *Clin Infect Dis* 1998; 27: 1442–50.
- Klugman KP, Koornhof HJ. Drug resistance patterns and serogroups or serotypes of pneumococcal isolates from cerebrospinal fluid or blood, 1979–1986. *J Infect Dis* 1988; 158: 956–64.

28. McGee L, Klugman KP, Tomasz A. Serotypes and clones of antibiotic-resistant pneumococci. In: Tomasz A, ed. *Streptococcus Pneumoniae: Molecular Biology and Mechanisms of Disease*, 1st edn. Larchmont: Mary Ann Liebert Inc., 2000: 375–9.
29. Jabes D, Nachnen S, Tomasz A. Penicillin-binding protein families: evidence for the clonal nature of penicillin resistance in clinical isolates of pneumococci. *J Infect Dis* 1989; 159: 16–25.
30. Laible G, Spratt BG, Hackenbeck R. Interspecies recombinational events during the evolution of altered PBP2X genes in penicillin-resistant clinical isolates of *Streptococcus pneumoniae*. *Mol Microbiol* 1991; 5: 1993–2002.
31. Coffey TJ, Dowson CG, Daniels M *et al.* Horizontal transfer of multiple penicillin-binding protein genes and capsular biosynthetic genes in natural populations of *Streptococcus pneumoniae*. *Mol Microbiol* 1991; 5: 2255–60.
32. Muñoz R, Coffey TJ, Daniels M *et al.* Intercontinental spread of multiresistant clone of serotype 23F *Streptococcus pneumoniae*. *J Infect Dis* 1991; 164: 302–6.
33. Carvalho C, Geslin P, Vaz Pato MV. Pulsed-field gel electrophoresis in *Streptococcus pneumoniae* isolated in France and Portugal. *Path Biol* 1996; 44: 430–4.
34. McDougal LK, Facklam R, Reeves M *et al.* Analysis of multiply antimicrobial resistant isolates of *Streptococcus pneumoniae* from the United States. *Antimicrob Agents Chemother* 1992; 36: 2176–84.
35. Klugman KP, Coffey TJ, Smith A *et al.* Cluster of an erythromycin-resistant variant of the Spanish multiply resistant 23F clone of *Streptococcus pneumoniae* in South Africa. *Eur J Clin Microbiol Infect Dis* 1994; 13: 171–4.
36. Lefèvre JC, Bertrand MA, Faucon G. Molecular analysis by pulsed-field gel electrophoresis of penicillin resistant *Streptococcus pneumoniae* from Toulouse, France. *Eur J Clin Microbiol Infect Dis* 1995; 14: 491–7.
37. Marchese A, Ramirez M, Schito GC, Tomasz A. Molecular epidemiology of penicillin-resistant *Streptococcus pneumoniae* isolates recovered in Italy from 1993 to 1996. *J Clin Microbiol* 1998; 36: 2944–9.
38. Sibold C, Wang J, Henrichsen J, Hakenbeck R. Genetic relationships of penicillin-susceptible and -resistant *Streptococcus pneumoniae* strains isolated on different continents. *Infect Immun* 1992; 60: 4119–26.
39. Yakupsky P, Porat N, Fraser D *et al.* Acquisition, carriage, and transmission of pneumococci with decreased antibiotic susceptibility in young children attending a day care facility in Southern Israel. *J Infect Dis* 1998; 177: 1003–12.
40. Setchanova L, Tomasz A. Molecular characterization of penicillin-resistant *Streptococcus pneumoniae* isolates from Bulgaria. *J Clin Microbiol* 1999; 37: 638–48.
41. Yoshida R, Hirakata Y, Kaku M *et al.* Trends of genetic relationship of serotype 23F penicillin-resistant *Streptococcus pneumoniae* in Japan. *Chemotherapy* 1997; 43: 232–8.
42. Davies T, Goering RV, Lovgren M *et al.* Molecular epidemiological survey of penicillin-resistant *Streptococcus pneumoniae* from Asia, Europe, and North America. *Diagn Microbiol Infect Dis* 1999; 34: 7–12.
43. Hall LMC, Whiley RA, Duke B *et al.* Genetic relatedness within and between serotypes of *Streptococcus pneumoniae* from the United Kingdom: analysis of multilocus enzyme electrophoresis, pulsed-field gel electrophoresis, and antimicrobial resistance patterns. *J Clin Microbiol* 1996; 34: 853–9.