

Review

Optical-Based (Bio) Sensing Systems Using Magnetic Nanoparticles

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Abstract: In recent years, various reports related to sensing application research have suggested that combining the synergistic impacts of optical, electrical or magnetic properties in a single technique can lead to a new multitasking platform. Owing to their unique features of the magnetic moment, biocompatibility, ease of surface modification, chemical stability, high surface area, high mass transference, magnetic nanoparticles have found a wide range of applications in various fields, especially in sensing systems. The present review is comprehensive information about magnetic nanoparticles utilized in the optical sensing platform, broadly categorized into four types: surface plasmon resonance (SPR), surface-enhanced Raman spectroscopy (SERS), fluorescence spectroscopy and near-infrared spectroscopy and imaging (NIRS) that are commonly used in various (bio) analytical applications. The review also includes some conclusions on the state of the art in this field and future aspects.

Keywords: optical sensor; magnetic nanoparticle; imaging; surface plasmon resonance; surface-enhanced Raman spectroscopy; fluorescence spectroscopy; near infrared spectroscopy

1. Introduction

In recent years, great interest has been shown in the field of nanotechnology in general and particularly its applications in the manufacturing of novel materials with added magnetic, electrical, optical, and biological properties for various areas. Among these nanomaterials with added properties, magnetic nanoparticles (MNPs) are quite interesting given their unique physico-chemical structure. The plethora of applications has been described for MNPs that are directly associated with modulation of the chemical structure, size distribution, and magnetic moment [1,2]. In addition, MNPs were found to possess a number of intrinsic properties such as biocompatibility, low toxicity, ease of surface modification, chemical stability, high surface area, high mass transference besides others [3]. Although all kind of MNPs, like other nanoparticles, can be used in different ways, size, and surface have a severe impact on the application area. For example, MNPs over 28 nm are ferrimagnetic and commonly used for magnetic separation and for electronic instruments as ferrofluids. MNPs below 28 nm, on the other side, are super-paramagnetic and are generally preferred for biomedical applications [4].

MNPs are generally divided, regarding their chemical structure, into metal oxides, pure metals, and magnetic nanocomposites. This distinct class of MNPs and application areas are depicted in

Figure 1. Metal oxides are most commonly used for biomedical applications, such as cell labeling and imaging [5–8], diagnostic and therapeutic agents [9]. Among metal oxides, iron oxides are commonly used since they have better biocompatibility [10]. In contrast to metal oxides, pure metals as a class of MNPs have limited application areas due to their chemical instability [11] and large size distributions [12]. Although rarely reported, the use of pure metal nanoparticles is generally found in electronic devices such as ultra-high-density magnetic storage devices [13–15]. In other respects, the use of magnetic nanocomposites, particularly in a sensor, biosensor and detection systems, has become increasingly common in recent years [16–19]. Disadvantages of MNPs, such as their tendency to aggregate to reduce surface energies, and oxidation in air, are among the major obstacles to the spread of their more frequent use in the applications [20]. Therefore, from the application point of view, researchers have proposed a new approach to the development of new materials by coating polymer, silica or other materials over MNPs, so-called magnetic nanocomposites [21–23].

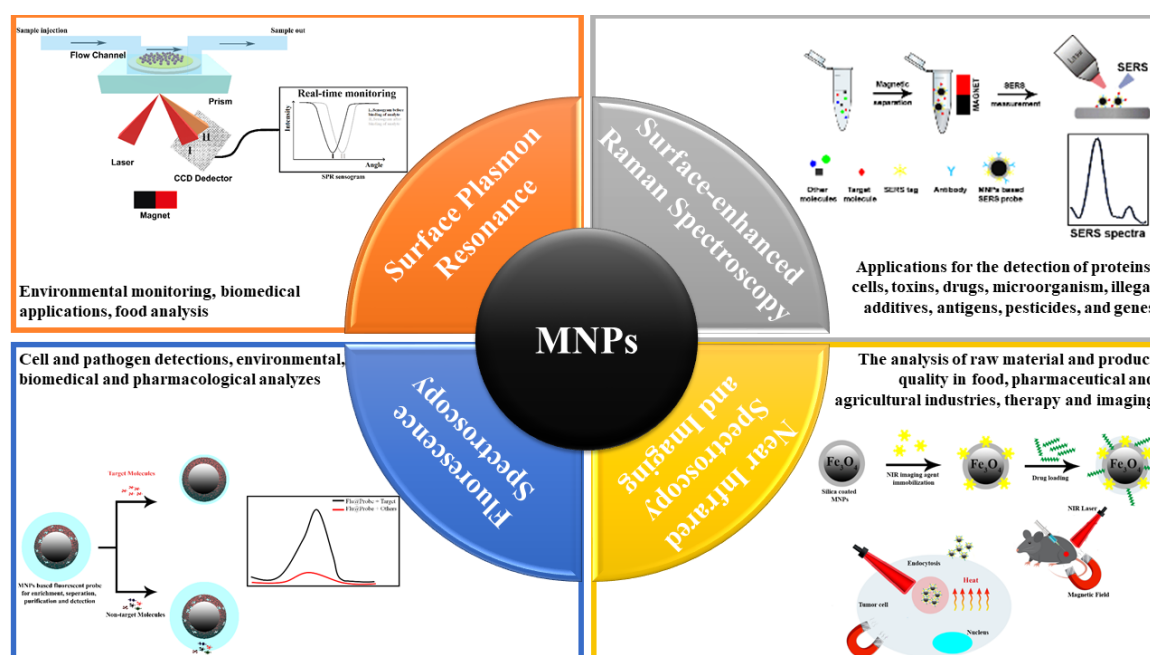


Figure 1. The main applications of optical-based (bio) sensing systems using magnetic nanoparticles (MNPs) in biomedical and environmental fields.

In practice, the feasible features of MNPs are not only affected by their structure but also by their fabrication techniques and surrounding environments [4,24,25]. MNPs can be prepared in the following three routes to suit the final desired application. The majority of these methods are based on the chemical methods or “bottom-up” synthesis procedures, which include co-precipitation, high-temperature thermal decomposition and/or reduction, sol-gel synthesis, flow-injection synthesis, oxidation method, electrochemical method, aerosol/vapor-phase method, supercritical fluid method, and synthesis using nanoreactors [5,25]. Chemical synthesis is controllable for almost all the processing conditions, and indeed, there has been a report indicating that this may be one reason for the “popularity” of chemical techniques for synthesis [26]. Alternatively, MNPs can be formed via physical methods through a simple non-chemical “top-down” dispersion technique, e.g., gas-phase deposition and electron-beam lithography [5,25,27]. Traditionally, physical methods have been the less preferred synthesis techniques for MNPs given an inability to control the size of particles down to the nanometer scale using these methods [27]. On the other hand, bio-reduction in the so-called microorganism approach, which has allowed large quantities of production, ensures the high yield, good reproducibility, and stability, as well as low cost [5,27]. In optical-based (bio)sensing systems, multifunctional composites such as $\text{Fe}_3\text{O}_4@\text{Ag}$, $\text{Fe}_3\text{O}_4@\text{Au}$, etc., are mainly prepared by

two methods: core-shell structure (1) and immobilization on the core (2) [28,29]. In both methods, magnetic nanoparticles are first coated with silica shells to increase the stability of the magnetic nanoparticles and to facilitate their modification. In the first of these directions, Ag or Au is reduced by using chemical compounds such as NaBH_4 , citric acid, etc. on the surface of the magnetic core. In the second method, gold or silver nanoparticles are prepared in different forms such as spherical, cubes or nanorods and then immobilized to the surface of the magnetic core by physical or chemical methods. In physical immobilization, it is usually carried out by modification of the magnetic core with polymers such as oleic acid, etc., and then bonded to the magnetic surface by the secondary interactions. In chemical immobilization, the surface of magnetic and Au or Ag nanoparticles are first modified with functional groups such as carboxyl or amine, and then the chemical immobilization is performed by modification method such as 1-Ethyl-3-(3-dimethylaminopropyl) carbodiimide/N-hydroxysuccinimide (EDC/NHS) coupling, etc.

After synthesis of MNPs, the common analytical techniques used for the characterization of MNPs are: ultraviolet (UV)–visible spectrophotometry, dynamic light scattering (DLS), scanning electron microscopy (SEM), transmission electron microscopy (TEM), Fourier transform infrared spectroscopy (FTIR), atomic force microscopy (AFM), powder X-ray diffraction (XRD), X-ray fluorescence (XRF) and energy-dispersive spectroscopy (EDS) [30,31].

There is no doubt that the extensive literature that can be associated with the term of biosensor shows that the related field of research is very attractive. In the first approach, a biosensor can be simply defined as a device that intimately combines a biological sensing element with a transducer [32]. In an efficient biosensor design, it is essential to convert the biological interaction to a signal which is transduced by physical, chemical, optical, thermal or electrochemical actions, into observable information and analyzed quantitatively [33]. Even though electrical and electrochemical platforms are the most advanced and used biosensor platforms, these systems have drawbacks owing to the extra reduction/oxidizing agent being involved [34]. While the biosensor can be built in a comparable manner based on optical and electrochemical transducers, it obviously demonstrates distinct working features [32]. Optical biosensors play a prominent role in sensing applications because they have distinct advantages: for example, high-precision detection and label-free design [34]. Optical biosensors also offer the possibility of immune electromagnetic interference, capable of remote sensing, and can perform multiplex detection within a single device [35]. The next generation of sensing equipment for daily use is optical biosensor platforms, based on absorbance, photoluminescence and surface plasmon resonance (SPR) [34]. In addition, coupling the synergistic effects of the optical properties and magnetic properties in a single method may offer new multitasking platforms for (bio)sensing, biolabeling/imaging, cell sorting/separation, and photothermal therapy [36].

In this comprehensive review, we will discuss optical biosensing systems based on the use of MNPs. Under this category, there exist a number of optical-sensing methods by using MNPs, including SPR, surface-enhanced Raman spectroscopy (SERS), fluorescence spectroscopy and near-infrared spectroscopy and imaging (NIRS).

2. Surface Plasmon Resonance (SPR)

Surface plasmon resonance (SPR) is a complex physical phenomenon occurring on an electrically conductive noble metal layer at the interface between two media (i.e. high refractive indices of sensor's glass surface and low refractive indices of buffer) when p-polarized light under conditions of total internal reflection [37,38]. Interest in the SPR effect started with an explanation as the energy of photons absorbed by the free electrons of the metallic layer reduced the amplitude of the incident light that comes with a certain angle. Despite the SPR effect being successfully explained in 1968, SPR as a method of determination in the field of biosensor was first mentioned in 1983 by Wijaya et al. Although fluorescence or absorbance based optical methods have been used since earlier times, SPR as surface-based optical method has been developed very rapidly and has attracted considerable interest in biosensing platform [38]. Investigation of biomolecular interactions with SPR is considered

an exceedingly powerful analysis method due to SPR's remarkable properties as an analytical tool such as its label-free nature, real-time, highly precise, a short time and simplicity. Despite the usefulness of SPR, the conventional SPR methods are mainly hampered by the detection of low concentration, low molecular weight biomolecules due to trivial changes of the refractive index in the binding process. Several approaches to overcome this challenge have been offered by researchers based on the use of nanoparticles [39,40]. Until now, decorating SPR surfaces with MNPs seems to be the most widely attested. Especially the application of MNPs in SPR sensing has attracted great focus owing to advantages which are (1) capture of the target molecules from the complex sample and (2) high refractive index and high molecular weight of MNPs to recruit the SPR signal [41,42]. Due to the aforementioned desired properties of MNPs, these compounds are widely used for environmental monitoring, biomedical applications, food analysis, etc. [43–46]. The principle of the SPR system with MNPs was illustrated in Figure 2 [47].

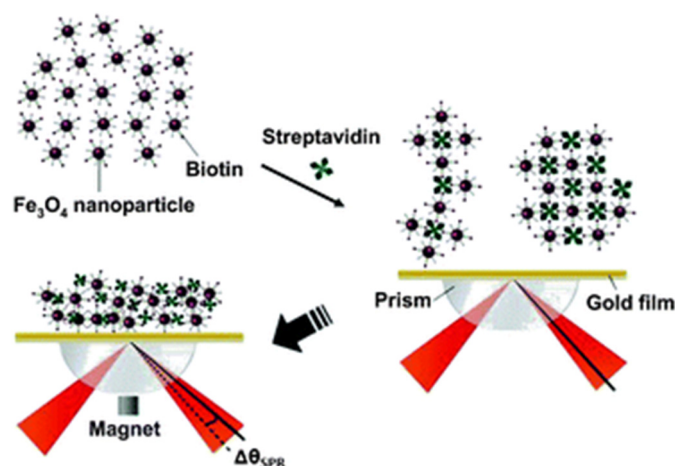


Figure 2. Amplification of the surface plasmon resonance (SPR) response signal with MNPs [47]. Reproduced with permission of the Royal Society of Chemistry.

In 2018, Sun and co-workers reported the SPR biosensor based on hollow gold nanospheres and improved sandwich assay with magnetic nanoparticles to detect rabbit IgG. While the reduced graphene layer (rGO) was covered with Ag nanoparticles to obtain SPR surface, Fe_3O_4 MNPs were utilized to capture the analyte of interest easily from the medium. In order to bind the antibody to the surface without modification and avoid agglomeration of nanoparticles, polydopamine (PDA) was used. These researchers showed that the detection of rabbit IgG by gold nanostructures resulted in the 132 times lower concentration when compared with a conventional SPR biosensor. The changes in the SPR signal caused by the magneto-sandwich immunoassay were further compared with the signal of hollow gold nanospheres. Their investigation showed that the 8 times higher enhancement in the SPR signal was related to the magneto-sandwich immunoassay [48]. In another study, aptamer immobilized SPR-based biosensors were developed by utilizing core-shell AuNPs and MNPs (MNPs) as the plasmonic coupling partners for the detection of thrombin as a model analyte. Aptamer immobilized AuNPs-MNPs conjugate-based SPR biosensors exhibited a considerable response to the thrombin with the limits of detection (LOD) of 0.6 nM. As compared to the SPR signal shift with a control group that is based on SPR signal without AuNPs-MNPs conjugate, AuNPs-MNPs conjugate based SPR angle shift was enlarged for 5 times at the 100 nM concentration [49]. A summary of the different applications of the MNPs by combining the synergistic effects of the SPR is summarized in Table 1.

Table 1. Examples of the application of MNPs in SPR system.

Magnetic Part	SPR System	Applications	Advantages	[R]
Active layer consisting of Fe ₃ O ₄ + Polyethylene glycol (PEG4000)	Silver-coated prism-coupled SPR System	Plant DNA	Target analyte could not be detected without using a magnetic part.	[50]
Polydopamine-Ag capped Fe ₃ O ₄ NPs reduced with graphene oxide	Gold film of SPR chip was electromagnetically coupled with hollow gold nanoparticles	Rabbit IgG	Detection limit 132 times lower than conventional SPR and 8 times lower than immunosandwich assay.	[48]
Gold capped Fe ₃ O ₄ nanoparticles (GMNPs)	Conventional SPR system with gold chip	Thrombin	SPR angle shift is enlarged for 5 times comparing with that of control group without GMNPs	[49]
MNPs conjugated with antibody	Conventional SPR system with gold chip	Pathogenic Bacteria	MNPs offer a sensing enhancement of 4 orders of magnitude.	[51]
Magneto plasmonic nanoparticles (core shell gold capped MNPs)	Conventional SPR system	Antigen (Tuberculosis marker protein)	Implementation of magneto-plasmonic NPs outcomes in 30-fold extension of the SPR signal at the limit of detection.	[52]
Nanohybrids containing Fe ₃ O ₄ NPs and hollow gold sphere nanoparticles.	SPR chip coated with carboxyl functionalized graphene oxide sheet.	Human IgG	Detection limit is approximately 260-fold lower than that acquired with sandwich assay by routine SPR biosensors.	[53]
Polydopamine-wrapped magnetic multi-walled carbon nanotubes	Capture antibody-immobilized SPR-sensing film modified by hollow gold nanoparticles and polydopamine	Protein (Human cardiac troponin I, CTnI)	Minimum detectable SPR response for the concentration of target molecule is 1000 times lower than that achieved by the traditional SPR immunoassay.	[54]
Magnetic fluid photonic crystal (i.e. kind of colloid compassed of MNPs dispersed in carrier liquid)	Kretschmann configuration (prism coupling) SPR system.	—————	The excellent benefit of elevated sensitivity is with the combination of magnetic fluid photonic crystal and SPR.	[55]
Aptamer-immobilized Fe ₃ O ₄ nanoparticles with organic clusters	Kretschmann configuration (prism coupling) SPR system.	Protein (prion disease associated isoform, PrP ^{Sc})	The SPR scheme involves magnetic NPs-organic clusters that allow for a 215-fold rise in the immediate SPR signal.	[46]
Streptavidin-coated MNPs	Grating-coupled SPR (GC-SPR) with wavelength interrogation.	Lipid (Extracellular vesicles, EVs)	Target could not be detected with the direct SPR detection platform at such low concentrations.	[56]
Fe-C core shell–aptamer conjugation	Prism-coupling SPR system	Protein (prion disease associated isoform, PrP ^{Sc})	The detection sensitivity of PrP ^{Sc} has been improved by about 10 times relative to the direct format of SPR detection.	[57]
Antibody-functionalized MNPs	Antibody-immobilized SPR immunoassay	Hormone (Estradiol)	The MNPs showed outstanding ability to amplify the SPR signal.	[45]
PEG-4000 functionalized Fe ₃ O ₄ MNPs	SPR set up device in Kretschmann configuration and He/Ne laser beam.	Microalgae	Microalgae population has been successfully monitored with the proposed system.	[58]
MNPs antibody conjugates	Conventional SPR spectroscopy	Cancer Biomarkers	To differentiate ovarian cancer, this multiplexed scheme accomplished sensitivity and specificity of up to 94% and 98%, respectively.	[59]

3. Surface-Enhanced Raman Spectroscopy (SERS)

Surface-enhanced Raman scattering (SERS) based on the enhancement of molecular Raman scattering by surface plasmons after the interaction between light and nanostructures is a surface-sensitive technique that is used for the detection of target with high molecular sensitivity and specificity at ultra-trace concentrations, including single molecules and provides more details on the chemical structure and conformation of target molecule [60]. The enhancement of the Raman signal is explained by two mechanisms: chemical enhancement arising from the charge transfer between the nanostructures and the adsorbed target and electromagnetic enhancement stemming from the electromagnetic field effect of the adsorbed target by stimulating the surface plasmons of nanostructures [61,62]. Since the enhancement of molecular Raman scattering mainly occurred by electromagnetic enhancement, novel nanostructures have been developed to achieve high enhancement factors by changing the plasmonic properties. The surface plasmons of nanostructures can be changed with the physical and chemical properties of nanostructures such as shape, size, type, and composition, etc. thus directly affecting the sensitivity and selectivity of SERS [63]. Because of their higher enhancement factors and the accessibility of plasmonic resonances in the visible and NIR areas, the plasmonic nanostructures (PNS) based on Au and Ag are most often preferred to produce SERS substrate [64]. Because of their higher enhancement factors and the accessibility of plasmonic resonances in the visible and NIR areas, the plasmonic nanostructures based on Au and Ag are most often preferred to produce SERS substrates. The SERS substrates have been successfully manufactured for the detection of targets such as biomolecules, cells, microorganisms, environmental pollutants, etc.

Although SERS has many important advantages, there are still needs for improvement of the reproducibility and stability of SERS substrates and the analysis of biological fluids and real environmental samples. These improvements can be achieved using MNPs or magnetic nanocomposites because of their unique properties such as strong superparamagnetic property, low toxicity, biocompatibility, easy preparation, and high adsorption ability. As outlined in Table 2, MNPs in SERS analysis have been used for the detection of proteins [65–68], cells [69–71], toxins [72], drugs [73,74], microorganisms [75–78], illegal additives [79,80], antigens [81,82], pesticides [83], and genes [84,85].

Table 2. Selected examples of surface-enhanced Raman spectroscopy (SERS) sensors based on MNPs.

Type of PNS	Size of PNS, nm	Forms of MNPs	Size of MNPs, nm	Reporter ¹	Analyte	Detection limit	[R]
Au NPs	15	γ -Fe ₂ O ₃ covered with silica shell and polymer shell	64	DTNB	Tau protein	25 fM	[65]
Au NPs	20	γ -Fe ₂ O ₃ MNPs covered with silica and Au shell	160	DTNB	Telomerase activity	1 cell/mL	[67]
Triangular Ag nanoprisms	40	γ -Fe ₂ O ₃ MNPs MNPs	21	MBA	Tumor cells	1 cell/mL	[69]
Au NPs	20	γ -Fe ₂ O ₃ covered with silica shell	25	MBA	Microcystin-LR	2.0pg/mL.	[72]
Au NPs	20	Fe ₃ O ₄ magnetic microspheres covered with SiO ₂ shell and Au nanoparticles	500	Cyanine	Sildenafil citrate	10 nM	[74]
Au NPs	31	γ -Fe ₂ O ₃ encapsulated with polymer	330	MBA and DSNB	<i>S. typhimurium</i>	10 cells/mL	[77]
Au NPs	30	γ -Fe ₂ O ₃ MNPs	500	DP	Chloramphenicol	1.0 pg/mL	[79]
Ag NPs	20	Fe ₃ O ₄ MNPs covered with GO and Ag NPs	500	-	Chloramphenicol	0.1 nM	[80]
Au NPs	60	ParamNPs	200	IR-792 and NB	West Nile virus Antigen and Rift Valley fever virus Antigen	5 fg/mL	[81]
Au NPs	30	γ -Fe ₂ O ₃ MNPs	500	DP	PSA antigen	5 pg/mL	[82]
Ag/SiO ₂ core-shell NPs	35	γ -Fe ₂ O ₃ MNPs covered with silica shell	50	Rhodamine B	DNA	5 μ M	[84]
Ag shell	30	γ -Fe ₂ O ₃ MNPs covered with Ag shell	300	Cyanine	miRNA	0.3 fM	[85]
Ag shell	8	γ -Fe ₂ O ₃ MNPs covered with Ag shell	300	PATP	Thiram	1.0 nM	[86]
Au@Ag core-shell NPs	32	γ -Fe ₂ O ₃ MNPs covered with polymer shell	36	MBT	Kanamycin	2 pg/mL	[87]
Ni@Au and Ni@Ag NPs	212 and 222	Ni MNPs	89	rhodamine 6G	rhodamine 6G	1 mM	[88]
Au Shell	35	γ -Fe ₂ O ₃ MNPs covered with Au shell	50	-	Microcystin-LR	3 fM	[89]
Au Shell	15	Fe ₃ O ₄ MNPs covered with Au shell	17-30	pthiocresol	Pthiocresol	4.5 pM	[90]
Au nanocubes and nanospheres	51	DNA modified Fe ₃ O ₄ magnetic beads	1000	Cyanine	DNA	1pM	[91]
Silver layers on reporter-coated AuNPs	60	Protein G modified Fe ₃ O ₄ magnetic beads	1000	MBA, DTNB, and TFMBA	Cytokines	4.5 pg mL	[92]

¹ DTNB: 5,5-dithiobis(2-dinitrobenzoic acid); DP: 4,4'-dipyridyl; MBA: 4-Mercaptobenzoic acid; NB: Infrared-792(IR-792) and Nile blue; PATB: p-aminothiophenol.

As mentioned above, gold (Au) nanostructures in the manufacture of SERS substrates are one of the most widely used metallic nanostructures for the plasmonic substrate. As an example, Zengin et al. [65] applied a sandwich assay for the ultrasensitive detection of tau protein by using monoclonal anti-tau immobilized hybrid MNPs as a probe and polyclonal anti-tau functionalized Au nanoparticles as SERS tags. Firstly, MNPs ($\gamma\text{-Fe}_2\text{O}_3$) were coated with silica for easy modification and then the hybrid structure was coated with polymer shell by Reversible Addition–Fragmentation chain Transfer (RAFT) polymerization. After the modification of the polymer shell, the monoclonal anti-tau was immobilized on the surface of the hybrid MNPs to obtain the capture probe. Gold nanoparticles were synthesized by citrate reduction with a diameter of 15 ± 8 nm and then, the gold nanoparticles were first modified with a layer of Raman reporter, 5,5-dithiobis(2-dinitrobenzoic acid) (DTNB) to obtain the homogeneous sandwich assay. Finally, their surfaces were functionalized with polyclonal anti-tau to prepare SERS tags. In this study, the MNPs used in sandwich assay provided easy and rapid detection of the tau protein with a detection limit of 25 fM. As another example, Yang et al. [79] performed a similar approach for the detection of chloramphenicol (CAP) by using magnetic separation. For the preparation of SERS tags, gold nanoparticles were first modified with 4,4'-dipyridyl (DP) as SERS reporter and then functionalized with chloramphenicol-bovine serum albumin (BSA) conjugate. CAP antibody was immobilized on the surface of carboxyl-functionalized magnetic beads (average diameter 500 nm) by EDC/NHS coupling reaction for magnetically and selectively separation of CAP. The limit of detection was obtained as 1.0 pg/mL for CAP in aqueous solution and also this SERS immunosensor system based on MNPs was applied to detect CAP in real samples and CAP was detected in the CAP concentration range of 0– 5×10^5 pg/mL. The selectivity experiment and real sample analysis indicate that this immunosensor is fast, sensitive and specific for the detection of CAP in the aqueous solution. As an example of toxin analysis, He et al. [72] developed an aptasensor based on SERS with MNPs for the detection of microcystin-LR (MC-LR) (Figure 3). After the synthesis of Au nanoparticles, its surface was modified with SERS reporter, MBA and then functionalized with MC-LR aptamer as SERS probe. MNPs were first covered with silica for easy immobilization of biomolecule and then the complementary DNA to MC-LR aptamer was immobilized on the surfaces of MNPs by biotin/avidin affinity as a capture probe. The developed aptasensor based on magnetic SERS was successfully applied to the selective and specific detection of MC-LR in tap water with the limit of detection, 2.0 pg/mL. As another approach, Yang et al. [81] developed an aptasensor based on magnetic SERS for the detection of prostate-specific antigen (PSA). PSA-aptamer was immobilized on the surface of the magnetic nanoparticle by EDC/NHS coupling reaction for the specific separation of PSA in the matrix. Au NPs were functionalized with DNA of PSA-complementary as a signal probe. The sandwich assay was applied for detection of PSA and LOD was obtained as 5.0 pg/mL and 25 pg/mL in aqueous solution and human serum, respectively. In another study for the detection of antigens, Neng et al. [82] performed the multiplex detection of antigens by using two sets of a spectrally distinct reporter for the preparation of the SERS probe. For the preparation of the SERS probe, the Raman reporter dyes Infrared-792 (IR-792) and Nile blue (NB) were bound separately on the surface of Au NPs and IR-792-coated Au NPs were modified with anti-E IgG and the NB-coated Au NPs were functionalized with anti-N IgG for specific recognition of each target antigen. For capture probe, paramagnetic nanoparticles were first coated with silica and then functionalized with anti-E and anti-N IgG. Briefly, the following procedure was applied for the assay: the antigens were first separated magnetically with the capture probe and then mixed with the antigen modified Au NPs. In the developed SERS sensor, LOD was around 5 fg/mL for both antigens. This study demonstrated the suitability of magnetic particles in SERS for the multiple detection of targets. In a study to examine the performance of SERS based on MNPs in the detection of microorganisms, polymeric MNPs were synthesized as a capture probe and Au NPs were modified with SERS reporter and specific antibody for the detection of *S. typhimurium* in the food product [77]. MBA (4-mercapto benzoic acid) and 5,5'-dithiobis(succinimidyl-2-nitrobenzoate) (DSNB) were used as SERS reporter. After the application of sandwich assay for the detection of microorganism, the limit of detection (LOD) of MBA and DSNB

was obtained as 100 cells/mL, and 10 cells/mL, respectively, in a spiked food product. The enhanced sensitivity of DSNB as a SERS reporter is due to its molecular characteristics that provide higher Raman scattering. In this study, higher reproducibility is obtained as it provides better solubility in water by coating the magnetic cores with polymer film.

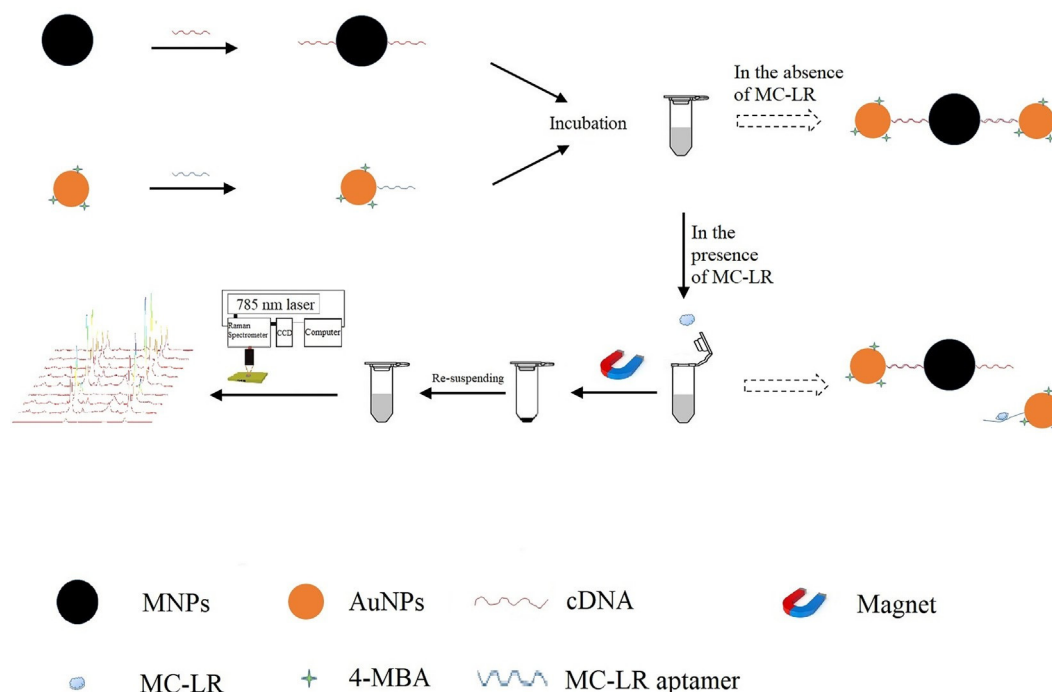


Figure 3. Illustration of the principle for the analysis of microcystin-LR (MC-LR) based on the SERS-based aptasensor. Reprinted [72] © 2019, with permission from Elsevier.

In SERS substrate, the other widely used nanostructures are silver nanostructures to create plasmonic surfaces. As an example, Liang et al. [84] developed a DNA hybridization assay based on magnetic SERS for DNA sequences related to HIV. In this strategy, Ag/SiO₂ core-shell nanoparticle-modified with Raman tags (Rhodamine B) were prepared as SERS probe and MNPs covered with silica shell were functionalized with oligonucleotides related HIV as capture probe. The hybridization reaction was performed between these probes. After the magnetical separation of DNA related to HIV from 3 μM solution, the DNA was successfully detected via the SERS probe. In another approach for the detection of genes, Pang et al. [85] developed a SERS sensor based on functionalized Fe₃O₄@Ag magnetic nanoparticle for the detection of microRNA (miRNA) in total RNA extract from cancer cells. In this application, unlike the others, MNPs were not only functionalized with the capture agent but also functionalized with the SERS signal reporter. This study has demonstrated the advantage of MNPs which are used to concentrate, capture and purify the target gene with the detection limit of 0.3 fM by using a single hybrid nanostructure. Furthermore, a similar strategy has been applied to the detection of microorganisms [76]. In another example, a seed-mediated strategy was applied to obtain an Ag shell on the surface of MNPs after the SiO₂ coating to use the detection of pesticide thiram [86] and the limit of detection was achieved as 1.0 nM for thiram.

The SERS applications of plasmonic nanostructures in different shapes are becoming quite attractive because of increasing the enhancement factor of SERS. As an example, Ruan et al. [69] developed SERS sensor based on triangular silver nanoprisms (AgNPR) and superparamagnetic iron oxide nanoparticles with a function of capture, enrichment, detection, and release for the tumor cells analysis. Firstly, AgNPR was modified with MBA and functionalized with rBSA (i.e., reductive bovine serum albumin) and FA (i.e., folic acid) generating the SERS probe. Then, MNPs were also modified with rBSA and FA generating the capture probe to be used for the capturing, enriching and detecting

of cancer cells from blood samples. The LOD was obtained as 1 cell/mL and the results demonstrate that nanoprisms have stronger electromagnetic enhancement compared to nanospheres due to their asymmetric shape. In another example of the SERS applications of plasmonic nanostructures in different shapes, the dual-mode nanoprobe based on SERS and fluorescence detection were developed by using Raman reporter-tagged Au@Ag core-shell nanorods and quantum dots onto the silica nanospheres for IgG detection as mentioned Figure 4 [66]. In this study, LOD was obtained as 0.1 pg/mL by applying a sandwich assay.

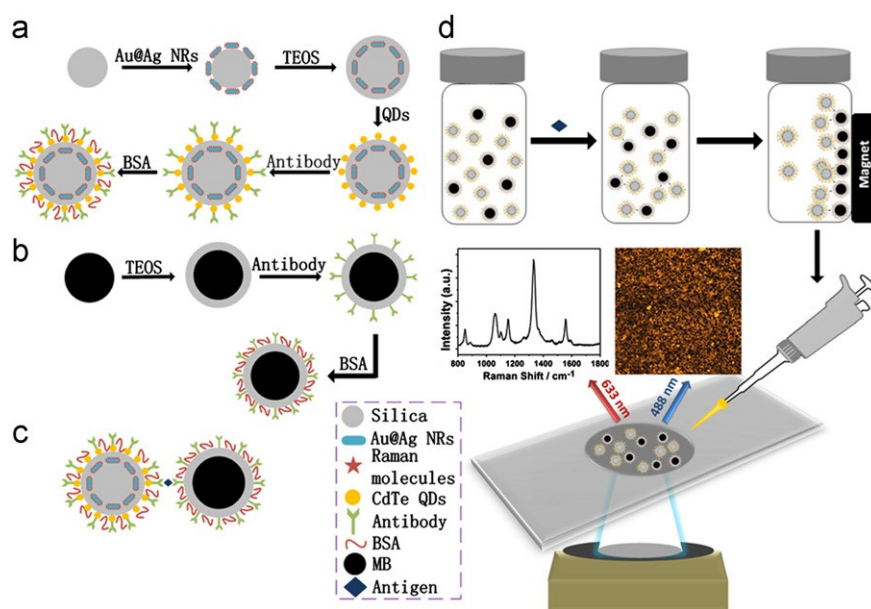


Figure 4. (a) Synthesis and surface functionalization procedure of the dual mode nanoprobe; (b) the synthesis and surface modification procedure of the magnetic beads (MBs); (c) illustration of the structure of the sandwich type nanocomplex formed during the immuno-reaction. For simplicity, only a single nanoprobe, antigen, and MB particle is drawn in the picture; (d) the aqueous phase immunoassay protocol. Reprinted [66] © 2013, with permission from Elsevier.

Zong et al. [67] developed a dual-mode sensor system based on colorimetry and magnetic SERS for the detection of telomerase. Au NPs were functionalized with DTNB and telomeric repeat complementary oligonucleotide to produce a bifunctional reporting tag in SERS and colorimetry. MNPs were first coated with silica and Au shell and then modified with telomerase substrate oligonucleotide as capture probe. The coating of Au shell provides easy modification and colorimetric detection via color change after the capturing. LODs of SERS and colorimetry were obtained as 1 tumor cell/mL and 10 tumor cells/mL, respectively.

As a consequence, MNPs are generally used in two different approaches in SERS applications. In the first one, it is designed as a capture probe for enrichment, purification and separation, and then the detection process by interacting with SERS tag based on the sandwich assay. In the other approach, the magnetic core is modified with SERS reporters such as BMA, DTNB, DP, etc. and plasmonic nanostructures such as Au or Ag shell and used directly as a capture probe and SERS probe. The approaches have advantages in itself as well as the approach being determined according to the analysis to be performed.

4. Fluorescence Spectroscopy

Various investigations have been conducted to address the impacts of the dual signal strategy to ameliorate the detection performance of the sensing system [93,94]. Different techniques that merge with MNPs have been used to serve as an all-in-one sensing system [95,96]. Among these

techniques, signal amplification labels relying on the enzyme or DNA labeling, dye labeling, and fluorophore labeling have exhibited great promise thanks to the synchronous properties [94,97,98]. Among numerous composite materials, the magneto-fluorescent nanocomposites have a great potential application due to their merits such as optical activity (biolabeling and bioimaging of fluorescent particles) and magnetic field guide opportunity [99–102]. Fluorometric assays have many advantages for biological applications such as higher sensitivity caused by their robust chemical and optical properties. Apart from their intrinsic properties, fluorescent particles have been frequently used for a sensing platform due to their outstanding advantages such as rapid, sensitive and low cost [103–105]. However, there are still some challenges such as an inevitable false signal caused by non-specific adsorption in real samples. Thus, the magnetization of particles was utilized to capture and separated target agents from complex matrices to avoid false signals [106–108]. Furthermore, MNPs can be adopted to utilize as contrast agents for magnetic resonance imaging (MRI) [100,109,110]. Magneto-fluorometric methods for sensing of numerous materials such as heavy metals [111,112], toxins [113], drugs [114,115], pesticide [116], biological materials i.e. DNA [117,118], cell [119,120], bacteria [121], etc. have been investigated. A summary of the different applications of fluorescence-based MNPs is given in Table 3.

Table 3. Some examples of fluorescence-based MNPs sensors.

Types of NPs	Capture probe	Size nm	Signal Probe	Size nm	Method	Target	LOD	[R]
MNPs@CuNCs	Folate receptor functionalized MNPs	300	Fluorescent copper nanoclusters	20	Sandwich assay	Streptavidin and biotin	0.47 nM and 3.1 nM	[93]
MNPs@FMs	Monoclonal antibodies (MAbs) immobilized Fe ₃ O ₄ MNPs	150	PABs functionalized fluorescent microspheres (FMs)	150	Microfluidic biosensor- smartphone based fluorescent microscopic system	Salmonella typhimurium	58 CFU/mL	[122]
MNP@SiO ₂ @BSA@Au-Myo-SNP@RhX	Fe ₃ O ₄ MNP@SiO ₂ @BSA@Au@antibody	292	SNP@RhX@antibody conjugates	85	Sandwich assay	myoglobin	0.28 ng/mL	[123]
MNPs-aptamer/TRFLNPs-cDNA	Fe ₃ O ₄ MNPs functionalized with ZEN aptamers	55	NaYF ₄ : Ce/Tb modified complementary DNA (TRFLNPs-cDNA)	34	Sandwich assay	Zearalenone (ZEN)	0.21 pg/mL	[124]
MNPs@ssDNA-FAM	Fe ₃ O ₄ MNPs functionalized with ssDNA	50	carboxyfluorescein (FAM)	-	Sandwich assay	Total mercury (Hg)	0.49 nM	[125]
Aptamer-conjugated FMNPs	Specific aptamer modified fluorescent magnetic nanoparticles (FMNPs)	100	-	-	Smartphone-based detection	Staphylococcus aureus	10 CFU/mL	[126]
CoFe ₂ O ₄ @dopamine@HSA@PDI-4NH ₂	CoFe ₂ O ₄ @dopamine@HSA@PDI-4NH ₂	190	PDI-4NH ₂ (Perylene diimide) carboxyfluorescein modified aptamer and dsDNA (FAMs@dsDNA)	-	Fluorescence imaging	Cell imaging	-	[127]
Fe ₃ O ₄ @aptamer 1@ IFN-γ@aptamer-FAMs@dsDNA	Fe ₃ O ₄ -aptamer 1/ IFN-γ/ aptamer	100-300	Morin	-	Aptamer/protein/aptamer/polymer supersandwich fluorescence sensor	Interferon gamma (IFN-γ)	0.175 fM	[128]
Fe ₃ O ₄ @SiO ₂ -NH ₂ -morin	Fe ₃ O ₄ @SiO ₂ -NH ₂	78	BSA-Au NCs	-	Fluorescence titrations	Cu ²⁺	7.5 nM	[129]
Fe ₃ O ₄ @SiO ₂ @Au MNPs	Fe ₃ O ₄ @SiO ₂ @Au MNPs	11	First target-specific DNA probe 1 (1pDNA) and bio-barcode DNA coated Au NPs (GNP)	-	H ₂ O ₂ quenching	glucose	3.0 μM	[130]
MNP-2nd DNA probe-target DNA-1st DNA probe-GNP-barcode DNA	Second target specific DNA probe modified Fe ₃ O ₄ MNPs	45	dye-labeled DNA	23	Sandwich assay	Exotoxin A gene sequence	1.2 ng/mL	[131]
MNPs-PEL-ssDNA	MNPs-PEL-ssDNA	78	-	-	Sandwich assay	lipopolysaccharide	35 ng/mL	[132]

MNPs@CuNCs: Magnetic nanoparticles-fluorescent copper nanoclusters; Rhodamine Red-X: RhX; SNP: Silica nanoparticle.

Liu et al. synthesized magnetic-fluorescent nanoparticles that were able to separate DNA. These research findings have indicated that the adsorption capacity was increased by using this approach. These nanoparticles also allowed to perform an experiment with only a small amount of nanocomposite except the necessity of radioactive probe [98].

According to the work of Cao et al., MNPs and highly fluorescent copper nanoclusters were prepared respectively by following approaches, streptavidin immobilization onto the surface afterward of the co-precipitation method and polymerization by using poly(T) as a template. The primary benefits of this research are that the dual-signal obtained from the supernatant and the re-dispersed precipitate confirmed each other and be integrated in order to result in a solid and quantitative detection [93].

In the case of Kim et al. research, dye-doped silica nanoparticles as fluorescent tagging and MNPs as a concentrating agent were utilized. The nanoparticles mentioned were used for the determination of enrofloxacin by combining with laser-induced fluorescence microscopy. The proposed technique has greatly improved the detection limit when it's compared with conventional methods such as ELISA [133].

A publication by Zarei-Ghobadi et al. has reported the fabrication of a genosensor for detection of target DNA based on the photoluminescence quenching of fluorescent dots in presence of Fe₃O₄ MNPs. The described nano(bio)sensor highlights the utilization of MNPs coated by the gold layer which enabled higher surface area as a quencher of fluorescent carbon dots. Also, the proposed sensor system was introduced as a good candidate for biosensing applications due to their advantages such as ease-synthesis, high biocompatibility, cost-effectivity and strong adsorption ability [134].

Another example of fluorescence shell tagging on MNPs was used to distinguish three cancer cell lines with different expression levels. A significant advantage in this study consists of the dual-mode nanoprobe which can be employed not only in the fluorescence measurements but also magnetic resonance-based cancer cell targeting [94].

Wang's group developed a bio-nanocomposite that has two parts i.e. superparamagnetic Fe₃O₄ nanoclusters as core and fluorescent dye molecule covalently doped onto silica layers to use an imaging and a photothermal therapy agent. In this contribution, bio-nanocomposite obtained were successfully applied in the specific photothermal therapy of colorectal cancer cells with high stability and reproducibility advantages [107].

Another genometric separation based on fluorescence assay was offered by Liu et al. for the detection of mercury in the canned fish sample. The proposed system was successfully applied for monitoring of trace amounts of mercury even after reused for 6 cycling times [125].

Niazi et al. were also developed a novel magneto fluorescent nanosphere by combining aptamer functionalized Fe₃O₄ and NaYF₄:Ce/Tb fluorescence nanoparticles for the detection of mycotoxin called zearalenone. Time-resolved fluorescence (TRFL) nanoparticles were labeled with complementary DNA to obtain the signal probe while amine-functionalized MNPs were labeled with zearalenone (ZEN) aptamer to obtain the capture probe. This novel bio-nanocomposite exhibits distinctive features such as stable, specific, convenient and rapid measurements. The schematic protocol for this magneto-fluorescent assay is shown in Figure 5 [124].

Fluorescent chemosensor assay was performed for the detection of target molecules which was based on internal charge transfer. Briefly, in the presence of target molecules interaction between magneto-fluorescent particles and Cu²⁺ that causes quenching due to the coordination with sensing platform was decreased distinctly because of the decomplexation result with an increase in the fluorescence intensity. The developed chemosensor in this study was successfully used for the monitoring of glutathione oxidized as a template molecule [135].

Another approach to use MNPs in combination with fluorescent structures has been proposed by Wang et al. Major novelty in this research was reported as utilizing not only fluorescent DNA nanotag but also enriched MNPs with DNA for the first time. Among various advantages, such as low non-specific adsorption, ultra-high sensitivity, easy preparation i.e., the proposed technique offers significant promise due to allowing the detection of even a single protein, in that case, human IgG [136].

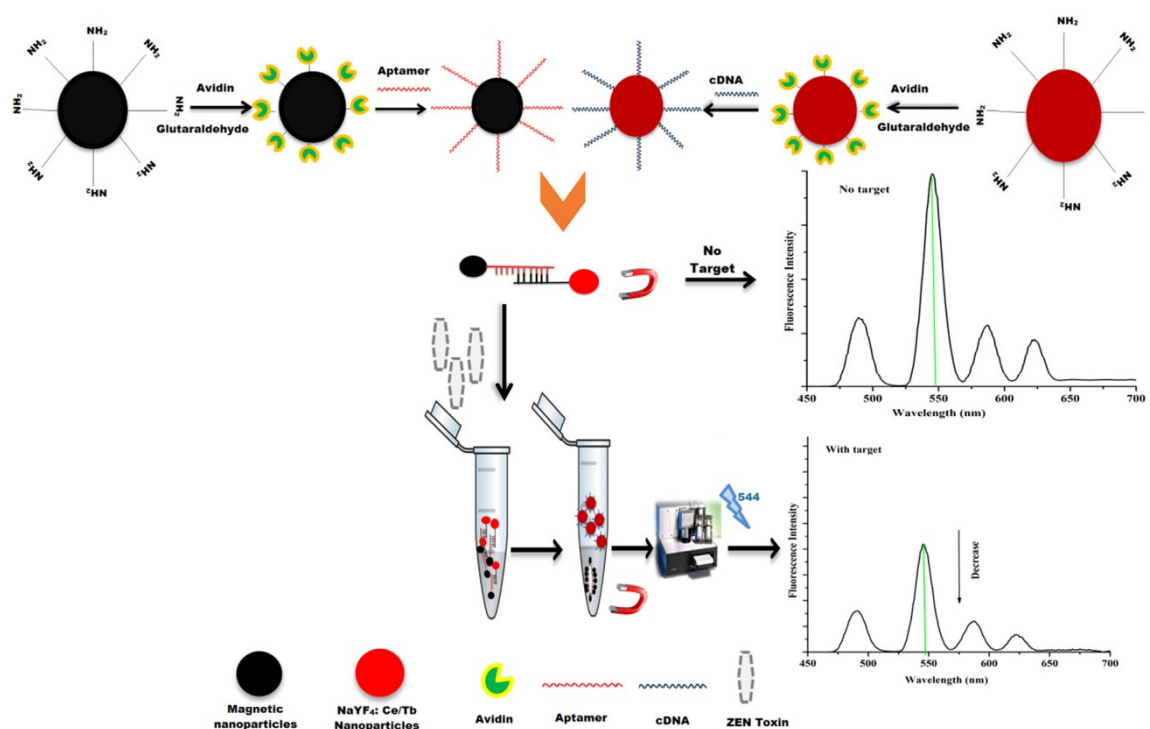


Figure 5. The detection principle based on MNPs as capture probe and fluorescent nanoparticles as a signal probe [124].

Wang and Jiang investigated solvothermal syntheses of $\text{CePO}_4\text{:Tb, Gd}$ hollow nanospheres which showed significant properties such as peroxidase mimetic activity and magnetic-fluorescent abilities into one nano-entity. The aforementioned properties made the prepared composite nanomaterials as potential applications in biocatalysis and bioimaging [110].

Other promising research was reported localized magnetic field distributions in a microchannel by using nickel powder [137]. They have captured the red fluorescent MNPs with the proposed platform [138–140]. Results have shown a great potential application in chemistry, biology, biomedicine and tissue engineering.

5. Near Infrared Spectroscopy (NIRS) and Imaging

Near-infrared spectroscopy (NIRS) based on the measurement of absorption of electromagnetic radiation, including wavelengths of 750 to 2500 nm is a type of vibration spectroscopy. The absorption bands originate mostly from the vibrations of molecular bonds such as C-H, O-H, C-O, and N-H within the structure [141]. The Spectra consisting of absorption bands allows the quantitative and qualitative analysis of the molecule being analyzed. However, the NIR radiation can easily penetrate into the body and also NIR radiation is scattered less than UV or visible radiation. Hence, the information based on imaging from the inner body can be obtained by analyzing its spectrum of NIRS [142]. Besides these features, NIRS has important advantages such as being fast and simple, and requiring little sample amount without pre-treatment. There are many applications for the analysis of raw material and product quality in food, pharmaceutical and agricultural industries [141,143,144]. Some applications are summarized in Table 4. In recent years, the new generation NIRS devices developed as functional near-infrared spectroscopy (fNIRS) have been also used for the diagnosis especially in biomedical applications such as neuroimaging, sugar blood, etc.

Table 4. Some applications of near-infrared spectroscopy (NIRS) sensors based on MNPs.

Types of NPs*	Size, nm	MNPs form	NIR agent	Target	Methods	Advantages	[R]
CMNPs	45	Fe ₃ O ₄	Cy5.5	Tumor cells	Specific cancer-targeting and magnetic resonance/near-infrared(MR/NIR) imaging	Dual imaging, excellent structural stability with biocompatible and biodegradable	[145]
Fe ₃ O ₄ @PPy-PEG	89	Fe ₃ O ₄	Polypyrrole (PPy)	Tumor cells	Photothermal therapy, magnetic targeting	Combination of combined the Fenton reaction and photothermal and magnet-guided cancer therapy	[146]
MFG-SiNc ₄	40	Magnetic graphene	SiNc ₄ and fluorescein	Tumor cells	Photodynamic/photothermal therapeutic	Time and cost effective treatments with a minimal therapy dose	[147]
DOX-Fe ₃ O ₄ @PAAP	170	Fe ₃ O ₄	Hydrophobic Fe ₃ O ₄	Doxorubicin (DOX)	In situ drug release and combined photothermal-chemotherapy	Fast and effective thermosensitive drug delivery	[148]
CdHgTe@DMF	100	Magnetic layered double hydroxide	Quantum dots of CdHgTe	Tumor cells	Drug delivery, optical bioimaging and magnetic targeted therapy	Slow-release curative effect and good cell imaging	[149]
Fe ₃ O ₄ -Aurods-Fe ₃ O ₄ nanodumbbells	70	Fe ₃ O ₄	Au Nanorods (Aurods)	Multiple pathogens	Detection, magnetic separation, and photokilling of multiple pathogens	Tunable nanoprobes for multiplex detection	[36]

* CMNPs: PSI-g-C12-PEG-folate cross-linked magnetic nanoparticles; MFG-SiNc₄: magnetic and fluorescent graphene (MFG) functionalized with SiNc₄; PAAP: poly (acrylamide-co-aniline)-g-PEG; DMF: dextran-magnetic layered double hydroxide-fluorouracil; PEG: Polyethylene glycole.

MNPs are widely used in NIRS due to their superior properties as targeting and MRI agent to improve the resolution and therapy by the development of dual-mode imaging and therapy systems. As an example, Zhou et al. [150] developed the multimodal theranostic platform by using the Fe₃O₄ magnetic nanoparticle as an MRI contrast agent and Indocyanine Green (IR820) as an NIR imaging agent. Firstly, Fe₃O₄-chitosan quaternary ammonium salt (CSQ-Fe) conjugate was synthesized to increase the stability and biocompatible of MNPs. Then, CSQ-Fe modified with IR820 and fluorescein isothiocyanate (FITC), separately. IR-CSQ-Fe and FITC-CSQ-Fe have similar size distribution and zeta potential and also their behavior in cellular uptake is quite comparable. Therefore, FITC-CSQ-Fe was used to determine the cellular uptake of IR820-CSQ-Fe. As a result, cell viability, long-time stability, MRI imaging, and photodynamic therapy tests indicate that IR-CSQ-Fe can be used as MRI and contrast agent for cancer diagnosis and detection. In another study, Zhou et al. [151] synthesized the Fe₃O₄ MNPs modified with NIR dye for sentinel lymph node mapping via the dual-mode imaging probe including MRI and NIR imaging. These conjugates performed better than small conventional dyes. Yang et al. [145] prepared biodegradable and biocompatible polymer-modified MNPs and functionalized with NIR dye Cy5.5 for specific cancer targeting and dual-mode imaging in vivo. Firstly, poly(succinimide) (PSI)-g-C12-PEG-folate was synthesized to improve biodegradable and biocompatible characteristics and then modified with Fe₃O₄ MNPs and NIR dye Cy5.5 for dual imaging. Consequently, MNPs were coated with a polymer and this hybrid material has an efficient NIR signal, high MRI sensitivity, an active targeting function, and good structural stability. Wu et al. [146] developed a new strategy for cancer imaging and therapy based on a combination of Fe₃O₄ MNPs and polypyrrole (PPy) in vivo Fenton reaction as illustrated in Figure 6. Fe₃O₄ magnetic core used for targeting the tumor site by applying the magnetic field was coated with PPy shell as near-infrared (NIR) light absorber for converting into heat for the photothermal ablation of tumors (PTT) and then modified functionalized with polyethylene glycol. Moreover, PPy coating not only provided PTT but also prevented Fe^{2+/3+} release from Fe₃O₄ MNPs. This approach indicated the first time that the releasing of iron ions from Fe₃O₄ in acidic medium and the Fenton reaction were enhanced by the photothermal effect. As a result, this hybrid nanoparticle obtained the combination of Fenton reaction and PPy as photothermal therapy provided efficient tumor damage without the effecting of normal organs. In another study, Huang et al. [152] synthesized chlorin e6 (Ce6) as a photosensitizer coated on the surface of Fe₃O₄ MNPs for gastric cancer imaging and therapy. As in other NIRS applications, MNPs have been used as targeting and MRI agents in this application and final multifunctional nanocarriers exhibited good water solubility and less cytotoxicity and good biocompatibility with high photodynamic

efficiency. As a consequence, the synthesized multifunctional nanocarriers performed targeting PDT and dual-mode NIR fluorescence imaging and MRI of gastric cancer tissue *in vivo*. In another similar study, graphene oxide was modified with ferrocene for targeting and functionalized with fluorescein for NIR fluorescence imaging [147]. After these modifications, the surface of nanoparticles was modified with silicon 2,3-naphthalocyanine bis (trihexylsilyloxi) (SiNc_4) as a photosensitizer. SiNc_4 was an ability to use in the photodynamic (PDT) and photothermal (PTT) therapies. This theranostic nanocarrier was used for cost and time-effective treatments with dual-mode imaging and phototherapy of cancer cells by a single light source. Cho et al. [153] designed the injectable nanocarrier based on hydrogel and collagen for drug delivery, phototherapy, and imaging. In this study, the high drug-loading capacity of hydrogel was utilized.

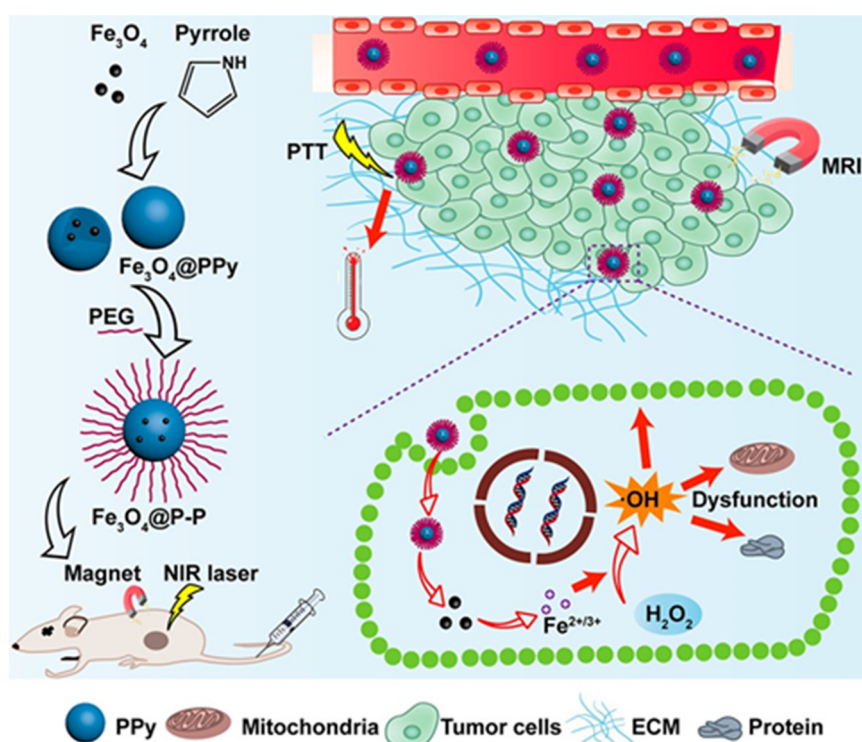


Figure 6. Schematic illustration of the synthetic route of Fe₃O₄@P-P NPs and the proposed synergistic antitumor mechanism of Fe₃O₄@P-P NPs. Reprinted with permission from [146]. Copyright American Chemical Society, 2018.

In addition to cell imaging and phototherapy, magnetic nanoparticle-mediated NIRS was used for pathogen detection. As an example, Wang and Irudayaraj [36] developed multifunctional nanoparticles based on gold nanorods and Fe₃O₄ MNPs for the simultaneous detection of multiple pathogens in a single sample, based on plasmon absorbance, magnetic separation, and thermal ablation. In this study, the plasmonic properties of gold nanorods were an ability to undertake multiple detections and MNPs were used for targeting, imaging and photothermal therapy.

Due to their small size and motility with the potential of the biomedical applications, microrobots have recently attracted attention. *In vivo* studies are especially prominent for the diagnosis and treatment of disease. In a study, Lee et al. [154] developed a thermosensitive hydrogel-based soft microrobot for targeting and drug delivery by applying NIR stimuli. The microrobot was mainly fabricated with alginate and NIPAM hydrogel for the thermosensitivity and drug loading and Fe₃O₄ MNPs were encapsulated into perfluoroalkoxy (PFA) tube for the targeting with a magnetic field. For drug release, a therapeutic drug, doxorubicin (DOX), was loaded into the microrobot. In this study, the drug delivery with the microrobot was performed efficiently in cancer cell treatment through *in vitro* Hep3B cell testing by NIR and electromagnetic actuation systems.

In conclusion, MNPs have been effectively used to develop multifunctional dual imaging and targeting systems for the purpose of drug release, targeting, thermal therapy, and imaging in NIRS applications.

6. Conclusions and Future Challenges

In recent years, MNPs have attracted much attention because of their unique properties such as biocompatibility, low toxicity, chemical stability, high surface area, easy synthesis, and modification, etc. to develop the multifunctional nanoparticle-based optical-sensing systems. Studies in this field generally consist of two main steps: (1) easy pre-modification process to improve low toxicity, solubility, and stability; (2) a second modification step with a specific recognition agent for targeting, imaging, diagnosis, separation, purification, and treatment. The most important disadvantage of MNPs is the aggregation and this problem can be overcome by the first modification such as silica or polymer coating, etc. The main disadvantage of optical sensors is the low sensitivity and selectivity of complex matrices such as body fluids, environmental samples, etc. and this problem is largely overcome by using the MNPs with a magnetic field. Moreover, the synthesis and characterization of MNPs need to be improved in (bio) sensing devices. In the future, optical sensors tend to miniaturization, implantability or wearability with high measuring accuracy for the multiple detections in (bio) sensing applications. Therefore, MNPs need to be adapted to such modifications for the integration of this field.

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