Spontaneous pregnancies in patients with at least one failed IVF cycle after the management of autoimmune disorders, hereditary thrombophilia, and methylation disorders

Atakan Tanacan¹, Mehmet Sinan Beksac¹

¹Division of Perinatology, Department of Obstetrics and Gynecology, Hacettepe University, Ankara, Turkey

ABSTRACT

Objective: This study aimed to describe the impact on achieving spontaneous pregnancy of treating patients with at least one failed in-vitro fertilization (IVF) cycle for autoimmune disorders, hereditary thrombophilia, and methylation disorders.

Methods: Fifty-three patients who met the enrollment criteria seen between January 2007 and October 2017 were included in this retrospective cohort study. The patients were retrospectively assessed for the presence of hereditary thrombophilia, methylenetetrahydrofolate reductase (MTHFR) polymorphisms, serum vitamin B12/folate/homocysteine levels, and autoimmune antibody positivity. The required data were extracted from the institutional patient database. Statistical analyses were performed on Statistical Package for the Social Sciences (SPSS.22[®]). The Kolmogorov-Smirnov test was used to evaluate the distribution of the data, and since the data did not following a normal distribution, proportions and median (minimum-maximum) values were used.

Results: The 53 patients included in the study had singleton pregnancies. The distribution of autoantibodies was as follows: thyroid peroxidase (n=17); antithyroglobulin (n=11); double-stranded DNA (n=4); antinuclear (n=8); anti-smooth muscle (n=1); and anticardiolipin IgG and IgM (n=1). Autoimmune diseases included Hashimoto's thyroiditis (n=23); SLE (n=7); Behcet's disease (n=1); Sjogren's syndrome (n=1); ulcerative colitis (n=1); and anti-phospholipid antibody syndrome (n=1). Ten patients had heterozygous Factor V Leiden thrombophilia; two had the prothrombin 20210A heterozygous mutation. Twenty-eight patients were positive for autoantibodies and hereditary thrombophilia and/or MTHFR polymorphisms.

Conclusions: Evaluation and management of hereditary thrombophilia, MTHFR gene polymorphisms, and/or autoimmune conditions may be beneficial for patients with unexplained infertility.

Keywords: infertility, *in vitro* fertilization, methylenetetrahydrofolate reductase, autoimmunity.

INTRODUCTION

Infertility is defined as a couple's inability to achieve pregnancy after abstaining from contraceptive methods for 12 months. The prevalence of primary infertility - a term coined to categorize couples unable to achieve their first pregnancy - is approximately 1.9% (Mascarenhas *et al.*, 2012). The time interval accepted for women aged 35 years or older is six months (Practice Committee of American Society for Reproductive Medicine, 2013).

In the absence of an explainable cause after comprehensive examination, couples are diagnosed with unexplained infertility (UI). Comprehensive examination is expected to reveal the following: 1) regular ovulation, 2) tubal patency, 3) a normal uterine cavity, 4) normal semen analysis results, and 5) an adequate ovarian oocyte reserve (Practice Committee of American Society for Reproductive Medicine, 2013). Expectant management, lifestyle changes, timed intercourse, and intrauterine insemination (IUI) with gonadotropin stimulation or *in vitro* fertilization (IVF) are different treatment options for UI (Nandi *et al.*, 2017; Quaas & Dokras, 2008).

IVF has been an effective treatment method for infertile couples performed with high success rates for nearly four decades (Steptoe & Edwards, 1978; Elizur *et al.*, 2006). It has become the preferred treatment modality for individuals with tubal factor infertility, diminished ovarian reserve, and severe male factor infertility (Rongieres, 2015). However, the role of IVF in patients with UI has not been clearly determined (Rongieres, 2015).

Maternal health disorders (epigenetic, metabolic, immunological conditions, etc.) that affect gametogenesis, fertilization, and implantation may also be linked to UI (Azem et al., 2004; Deroux et al., 2017; Coulam & Jeyendran, 2009; Zhong et al., 2012; Qublan et al., 2006). It has been reported that metabolic pathway disorders including aminoacyl-tRNA biosynthesis, "glycine-ser-ine-threonine" metabolism, "alanine-aspartate-glutamate" metabolism, "valine-leucine-isoleucine" biosynthesis, and methionine metabolism may be linked to impaired gametogenesis and/or unsuccessful implantation (Spiridonov et al., 2005; Ni et al., 2016; Unni et al., 2015). Autoimmune disorders (autoimmune antibody positivity and related diseases) may also be responsible for undesired biological processes such as uterine inflammation, which further lead to impaired endometrial receptivity and placentation (Deroux et al., 2017; Chen et al., 2017). Thus, methylation disorders and immunological conditions seem to play important roles in the pathophysiology of UI (Azem et al., 2004; Coulam & Jeyendran, 2009).

This study aimed to describe the impact on achieving spontaneous pregnancy of treating patients with at least one failed in-vitro fertilization (IVF) cycle for autoimmune disorders, hereditary thrombophilia, and methylation disorders.

MATERIALS AND METHODS

We evaluated 53 patients with at least one failed IVF cycle for the treatment of primary UI who became pregnant spontaneously and gave birth at our hospital (between January 2007-October 2017) as a consequence of proper management of their medical conditions (autoimmune disorders, hereditary thrombophilia, and MTHFR polymorphisms). The required data were extracted from the Hacettepe University patient database.

All patients with primary UI and at least one failed IVF cycle were included in a special follow-up program and evaluated before conception. The patients were screened

for hereditary thrombophilias (factor V Leiden mutation, prothrombin 20210A mutation, protein C/S deficiency), MTHFR polymorphisms, serum vitamin B12/folate/homocysteine levels, autoimmune antibody positivity (antinuclear (ANA), thyroid peroxidase (TPO), antithyroglobulin (TgAb), anti-parietal cell (APA), and antiphospholipid antibodies depending on their clinical findings and family history), and related disease/syndromes at the Hacettepe University, Division of Perinatology. Patients in a quiescent period in terms of immunological and metabolic inflammatory processes were allowed to get pregnant.

All patients were included in a special antenatal care program and had low molecular weight heparin (LMWH = enoxaparin 2000 Anti-Xa IU/0.2 ml), oral prednisone (methylprednisolone 4 mg), and aspirin (acetylsalicylic acid 100 mg) added to their specific treatment protocols as soon as they became pregnant. A special low-methionine diet was prescribed to patients with MTHFR polymorphisms.

Pregnancy follow-up comprised serial laboratory testing and ultrasound examination, aneuploidy screening (combined or triple test), fetal anatomy scanning at the 20th-24th week of gestation, oral glucose challenge test, and non-stress tests weekly (after the 28th week of gestation). All medications were stopped three days before delivery.

The median (minimum-maximum) values of patient age; duration of infertility; number of previous IVF cycles; serum folate, vitamin B12, and homocysteine levels just before conception; gestational week at birth; birth weight; and 5-minute APGAR scores were calculated, as the data did not follow a normal distribution. The proportions of patients with positive tests for autoantibodies, autoimmune diseases, hereditary thrombophilia, and MTHFR polymorphisms were also recorded.

Statistical analyses were performed on the Statistical Package for the Social Sciences (SPSS.22[®], IBM SPSS Statistics for Windows, Version 22.0. Armonk, NY: IBM Corp.). The Kolmogorov-Smirnov test was used to assess the distribution of the data, and since the data did not follow a normal distribution, proportions and median (minimum-maximum) values were used.

This study was approved by the Ethics Committe at Hacettepe University Et (GO 18/158).

RESULTS

The 53 singleton pregnancies resulted in live births. There were no ectopic pregnancies, miscarriages, or stillbirths. There were only three spontaneous late preterm births (5.6%) (at 34, 35, and 35 weeks of gestation). Preeclampsia, eclampsia, and placental abruption were not observed in any of the patients.

Table 1 shows the median and minimum-maximum values for maternal age; number of previous IVF cycles; duration of infertility; gestational week at birth; birth weight; 5-minute APGAR score; and serum folate, vitamin B12, and homocysteine levels.

Table 2 shows the proportions for autoantibodies. Table 3 shows the proportions for hereditary thrombophilias, MTHFR polymorphisms, and autoantibody positivity together with thrombophilias.

Twenty-three patients (43.4%) had Hashimoto's thyroiditis; seven (13.2%) had SLE; one (1.9%) had Behcet's Disease; one had Sjogren's syndrome; one (1.9%) had ulcerative colitis; and one (1.9%) had anti-phospholipid antibody syndrome.

The 53 patients had at least one hereditary thrombophilia and/or MTHFR polymorphism. Autoantibody positivity was seen in 28 patients (52.8%) combined with hereditary thrombophilia and/or MTHFR polymorphisms.

DISCUSSION

Although IVF is the most successful treatment method for UI, its overall live birth rate is approximately 40%, depending on maternal age and the number of transferred embryos (Elizur *et al.*, 2006). Infertility is a multifactorial condition that affects couples in medical, psychological, sociological, and economic aspects (Péloquin *et al.*, 2018). Some identifiable risk factors are genetic abnormalities, ovulatory disorders, tubal damage, uterine or peritoneal problems, and male factors (Ray *et al.*, 2012). On the other hand, nearly 30% of couples have no determined etiology (Ray *et al.*, 2012).

The complexity of the situation emphasizes the need for further investigation. Hyperhomocysteinemia, MTHFR gene polymorphisms, variations in folate pathway genes, vitamin B-complex deficiencies, hereditary thrombophilias, and autoimmune antibody production are some factors that contribute to the etiology of UI (Deroux *et al.*, 2017; Altmäe *et al.*, 2010; Das *et al.*, 2015; Hou *et al.*, 2016; Enciso *et al.*, 2016).

Thrombophilia can cause infertility as it impairs oocyte quality, increases hypoxic damage, and diminishes vascularization (Qublan et al., 2006). Properly vascularized and oxygenated follicles lead to higher rates of implantation (Qublan et al., 2006; Chui et al., 1997). It has been shown that during IVF the mean follicular diameter, oocyte retrieval rate, number of mature oocytes, and fertilization rates are higher in highly vascularized follicles (Qublan et al., 2006; Bhal et al., 1999). Implantation, placentation, angiogenesis, and vascular remodeling require an appropriate balance between coagulation and fibrinolysis, although thrombosis in the intervillous space and placental vessels may cause placental insufficiency (Qublan et al., 2006). On the other hand, heparin has a role in trophoblast differentiation and invasion (Lodigiani et al., 2017; Nelson & Greer, 2008). More specifically, low-molecular-weight heparin (LMWH) affects matrix metalloproteinases (MMP), tissue inhibitors, cadherin-E, heparin-binding epidermal growth factor (HBEGF), and insulin-like growth factor (IGF) (Lodigiani et al., 2017; Nelson & Greer, 2008; Di Simone et al., 2007; Erden et al., 2006; Das et al., 1994; Lacey et al. 2002). LMWH also has a positive effect on IVF outcomes (Lodigiani et al., 2017). These findings support our results, which indicated that appropriate management of thrombophilia and proper therapy with heparin play a significant role in the treatment of UI.

Methionine is an essential amino acid in nucleic acid synthesis and homocysteine is a metabolite resulting from its metabolism (Das et al., 2015). High levels of homocysteine may arise from deficiency of certain vitamins (folic acid, vitamin B6, etc.) and/or MTHFR gene polymorphisms (Das et al., 2015). Hyperhomocysteinemia causes many adverse pregnancy outcomes, including pregnancy loss, neural tube defects, chromosomal aneuploidies, fetal cardiac defects, preeclampsia, placental abruption, and intrauterine growth restriction (IUGR) (Das et al., 2015; Ray & Laskin, 1999; Botto & Yang, 2000). Hyperhomocysteinemia has also been associated with impaired follicular development, oxidative damage to oocytes, improper vascularization of the chorionic villi, and implantation failure (Das et al., 2015; Jerzak et al., 2003). Therefore, deficiencies in folate and/or B-complex vitamins lead to the accumulation of homocysteine and may cause UI (Altmäe et al., 2010). Some of the hypothesized mechanisms include impaired cell division, increased production rates of inflammatory cytokines, impaired nitric oxide metabolism, increased oxidative stress, increased rates of apoptosis, and impaired methylation reactions. All these factors combined affect oocyte development and embryo implantation **Table 1.** Descriptive statistics for maternal age; number of previous IVF cycles; duration of infertility; gestational week at birth; birth weight of the infants; 5-minute APGAR score; and serum levels of folate, vitamin B12, and homocysteine

Variables	Median	Minimum-Maximum
Maternal Age (years)	30	21-41
Number of previous IVF cycles	2	1-6
Duration of infertility (months)	24	12-72
Gestational week at birth	37	34-39
Birth weight of the infants (g)	3030	2100-3780
Fifth minute APGAR Score	10	7-10
Serum folate levels (nmol/L)	16.3	5.7-27
Serum vitamin B12 levels (ng/L)	289	129-465
Serum homocysteine levels (µmol/L)	8.1	3.4-12.4

Table 2. Percentage of autoantibodies		
Autoantibody	Percentages	
anti-TPO	17/53 (32.1%)	
anti-TG	11/53 (20.7%)	
ANA	8/53 (15.1%)	
Anti-dsDNA	4/53 (7.5%)	
ASMA	1/53 (1.9%)	
aCL IgG-IgM	1/53 (1.9%)	

anti-TPO: anti- thyroid peroxidase, anti-TG: antithyroglobulin, ANA: antinuclear antibodies, anti-dsDNA: anti-double stranded DNA antibody, ASMA: anti-smooth muscle antibody, aCL: anticardiolipin antibody.

Table 3. Proportion of hereditary thrombophilias, MTHFR polymorphisms, and autoantibody positivity together with thrombophilias		
Variables	Proportion	
MTHFR C677T Heterozygous	29/53 (54.7%)	
MTHFR C677T Homozygous	9/53 (17.0%)	
MTHFR A1298C Heterozygous	28/53 (52.8%)	
MTHFR A1298C Homozygous	8/53 (15.1%)	
MTHFR Compound Heterozygous	21/53 (39.6%)	
F 5 Leiden Heterozygous	10/53 (18.9%)	
F 5 Leiden Homozygous	2/53 (3.8%)	
Prothrombin 20210 A Heterozygous	2/53 (3.8%)	
Prothrombin 20210 A Homozygous	0/53 (0.0%)	
PAI-1 4G/5G Heterozygous	11/53 (20.7%)	
PAI-1 4G/5G homozygous	2/53 (3.8%)	
Autoantibody positivity with thrombophilia	28/53 (52.8%)	

MTHFR: Methylene tetrahydrofolate reductase, PAI: Plasminogen activator inhibitor

and reduce endometrial receptivity (Altmäe *et al.*, 2010). Thus, supplementation with folate and B-complex vitamins before conception decreases homocysteine levels in the follicular fluid, which leads to decreased rates of infertility and lower rates of miscarriage (Altmäe *et al.*, 2010; Chavarro *et al.*, 2008). Heterozygous mutations of the gene coding the MTHFR enzyme decrease enzyme activity by 35%, while homozygous mutations decrease activity by 70%, thus causing impaired folate metabolism

and accumulation of homocysteine (Frosst *et al.*, 1995). Several studies in the literature have described the negative effects of high homocysteine levels in patients with infertility and/or polycystic ovary syndrome (PCOS) (D'Uva *et al.*, 2007; Bibi *et al.*, 2010; Schachter *et al.*, 2003). These findings are consistent with our results. Appropriate management of decreased serum homocysteine levels and increasing serum folate and vitamin B levels are essential for the treatment of UI.

Autoimmune factors seem to play a pivotal role in infertility (Chen et al., 2017). The prevalence of antiphospholipid antibodies (aPL), ANA, and antithyroid antibodies was found to be particularly higher in women with UI (Chen et al., 2017). Implantation is a process that necessitates immunologic tolerance and requires cross-talk between the embryo and the maternal immune system (Kushnir et al., 2016). Impaired implantation and high pregnancy loss rates in women with autoimmune antibodies may be caused by injury to the syncytiotrophoblasts, endovascular trophoblasts covering the tip of the spiral arteries, endothelial cells of the spiral veins, superficial/glandular epithelial cells of the decidua, autoantibody inflammatory processes, and entrance of cell degradants from these tissues into the maternal circulation. These biological events result in impaired implantation and disturbed fetal perfusion (Beksac et al., 2017). In addition to these, ovulation, implantation, and onset of labor are thought to be associated with inflammation (Kushnir et al., 2016; Weiss et al., 2009). Although exact screening and management protocols for infertile patients positive for autoimmune antibodies have not been established, it seems reasonable to devise a patient-centered, individualized approach for these patients. Low-dose methylprednisolone (4 mg, once a day), low-dose acetylsalicylic acid (100 mg once a day), and low-dose LMWH (20 mg/0.2 ml enoxaparin sodium administered intramuscularly once a day) were the main components of treatment, as it has been shown that heparin and heparin-related derivatives have an anti-inflammatory effect (Young, 2008).

Although IVF is a revolutionary treatment for infertile couples, use without appropriate indication may increase both costs and procedure-related complications (Ben-Ami *et al.*, 2017). Ovarian hyperstimulation syndrome, multiple pregnancy, preterm labor, preeclampsia, and increased rates of chromosomal/structural abnormalities are some of the complications related with the IVF procedure (Ben-Ami *et al.*, 2017; Xu *et al.*, 2015).

The 53 patients had at least one underlying condition (hereditary thrombophilia, MTHFR gene polymorphism, or autoantibody positivity). After the introduction of appropriate management protocols, all patients delivered healthy babies. Preeclampsia, eclampsia, and placental abruption were not observed in any of the patients. There were only three spontaneous late preterm deliveries (5.6%) (at 34, 35 and 35 weeks of pregnancy, respectively).

The limitations of this study are the small number of patients, its retrospective design, and the fact that it reflects the experience of a single center. Furthermore, this is a descriptive assessment of a very specific group of patients who gave birth at our institution without the aid of assisted reproductive technology and who had a history of failed IVF cycles for UI. This study was not designed to investigate the success rate of our management protocol, but to stress the importance of knowing the etiological factors behind UI.

In conclusion, infertility is a field with plenty to discover. Underlying maternal conditions may cause conception failure. Evaluation and management of hereditary thrombophilias, MTHFR gene polymorphisms, and/or autoimmune conditions may be beneficial for primary infertility patients with at least one failed IVF cycle.

CONFLICT OF INTEREST

The authors have no conflict of interest to declare.

Funding/Support Statement

No funding was used fort his study.

ACKNOWLEDGMENTS

The authors would like to thank the medical staff of our institution for working tirelessly to provide their patients with optimal healthcare.

Corresponding author:

Atakan Tanacan Hacettepe University Hospital Division of Perinatology, Department of Obstetrics and Gynecology Sihhiye/Ankara E-mail: atakantanacan@yahoo.com

REFERENCES

Altmäe S, Stavreus-Evers A, Ruiz JR, Laanpere M, Syvänen T, Yngve A, Salumets A, Nilsson TK. Variations in folate pathway genes are associated with unexplained female infertility. Fertil Steril. 2010;94:130-7. PMID: 19324355 DOI: 10.1016/j.fertnstert.2009.02.025

Azem F, Many A, Yovel I, Amit A, Lessing JB, Kupferminc MJ. Increased rates of thrombophilia in women with repeated IVF failures. Hum Reprod. 2004;19:368-70. PMID: 14747183 DOI: 10.1093/humrep/deh069

Beksaç K, Örgül G, Çagan M, Karaağaoğlu E, Arslan S, Beksaç MS. Retrospective evaluation of pregnant women with celiac disease. J Turk Ger Gynecol Assoc. 2017;18:56-9. PMID: 28506952 DOI: 10.4274/jtgga.2016.0198

Ben-Ami I, Raziel A, Gidoni Y, Fauser BC, Ron-El R. Indications for in vitro fertilization treatment: From diagnosis to prognosis. In: Gardner DK, Weissman A, Howles CM, Shoham Z, eds. Textbook of Assisted Reproductive Techniques Volume 2: Clinical Perspectives. 5th ed. Boca Raton: CRC Press; 2017. p. 80-94.

Bhal PS, Pugh ND, Chui DK, Gregory L, Walker SM, Shaw RW. The use of transvaginal power Doppler ultrasonography to evaluate the relationship between perifollicular vascularity and outcome in in-vitro fertilization treatment cycles. Hum Reprod. 1999;14:939-45. PMID: 10221224 DOI: 10.1093/humrep/14.4.939

Bibi S, Pir MA, Qazi RA, Qureshi MB. Hyperhomocysteinemia in Pakistani women suffering from unexplained subfertility. Int J Reprod Med. 2010;8:76-9.

Botto LD, Yang Q. 5,10-Methylenetetrahydrofolate reductase gene variants and congenital anomalies: a HuGE review. Am J Epidemiol. 2000;151:862-77. PMID: 10791559 DOI: 10.1093/oxfordjournals.aje.a010290

Chavarro JE, Rich-Edwards JW, Rosner BA, Willett WC. Use of multivitamins, intake of B vitamins, and risk of ovulatory infertility. Fertil Steril. 2008;89:668-76. PMID: 17624345 DOI: 10.1016/j.fertnstert.2007.03.089

Chen X, Mo ML, Huang CY, Diao LH, Li GG, Li YY, Lerner A, Shoenfeld Y, Zeng Y. Association of serum autoantibodies with pregnancy outcome of patients undergoing first IVF/ICSI treatment: A prospective cohort study. J Reprod Immunol. 2017;122:14-20. PMID: 28804023 DOI: 10.1016/j.jri.2017.08.002 Coulam CB, Jeyendran RS. Thrombophilic gene polymorphisms are risk factors for unexplained infertility. Fertil Steril. 2009;91:1516-7. PMID: 18930220 DOI: 10.1016/j.fertnstert.2008.07.1782

gramme: a preliminary study. Hum Reprod. 1997;12:191-6.

PMID: 9043927 DOI: 10.1093/humrep/12.1.191

Das SK, Wang XN, Paria BC, Damm D, Abraham JA, Klagsbrun M, Andrews GK, Dey SK. Heparin-binding EGF-like growth factor gene is induced in the mouse uterus temporally by the blastocyst solely at the site of its apposition: a possible ligand for interaction with blastocyst EGF-receptor in implantation. Development. 1994;120:1071-83. PMID: 8026321

Das V, Misra D, Agrawal S, Agrawal A, Pandey A. Hyperhomocysteinemia and MTHFR gene 677 C>T polymorphism: questionable role in female infertility. Int J Reprod Contracept Obstet Gynecol. 2015;4:683-9. DOI: 10.18203/2320-1770.ijrcog20150074

Deroux A, Dumestre-Perard C, Dunand-Faure C, Bouillet L, Hoffmann P. Female infertility and serum auto-antibodies: a systematic review. Clin Rev Allergy Immunol. 2017;53:78-86. PMID: 27628237 DOI: 10.1007/s12016-016-8586-z

Di Simone N, Di Nicuolo F, Sanguinetti M, Ferrazzani S, D'Alessio MC, Castellani R, Bompiani A, Caruso A. Low-molecular weight heparin induces in vitro trophoblast invasiveness: role of matrix metalloproteinases and tissue inhibitors. Placenta. 2007;28:298-304. PMID: 16750851 DOI: 10.1016/j.placenta.2006.04.001

D'Uva M, Di Micco P, Strina I, Alviggi C, Iannuzzo M, Ranieri A, Mollo A, De Placido G. Hyperhomocysteinemia in women with unexplained sterility or recurrent early pregnancy loss from Southern Italy: a preliminary report. Thromb J. 2007;5:10. PMID: 17625006 DOI: 10.1186/1477-9560-5-10

Elizur SE, Lerner-Geva L, Levron J, Shulman A, Bider D, Dor J. Cumulative live birth rate following in vitro fertilization: study of 5,310 cycles. Gynecol Endocrinol. 2006;22:25-30. PMID: 16522530 DOI: 10.1080/09513590500453916

Enciso M, Sarasa J, Xanthopoulou L, Bristow S, Bowles M, Fragouli E, Delhanty J, Wells D. Polymorphisms in the MTH-FR gene influence embryo viability and the incidence of aneuploidy. Hum Genet. 2016;135:555-68. PMID: 27068821 DOI: 10.1007/s00439-016-1652-z

Erden O, Imir A, Guvenal T, Muslehiddinoglu A, Arici S, Cetin M, Cetin A. Investigation of the effects of heparin and low molecular weight heparin on E-cadherin and laminin expression in rat pregnancy by immunohistochemistry. Hum Reprod. 2006;21:3014-8. PMID: 16997938 DOI: 10.1093/humrep/del262

Frosst P, Blom HJ, Milos R, Goyette P, Sheppard CA, Matthews RG, Boers GJ, den Heijer M, Kluijtmans LA, van den Heuve LP, Rozen R. A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase. Nat Genet. 1995;10:111-3. PMID: 7647779 DOI: 10.1038/ng0595-111 Hou N, Chen S, Chen F, Jiang M, Zhang J, Yang Y, Zhu B, Bai X, Hu Y, Huang H, Xu C. Association between premature ovarian failure, polymorphisms in MTHFR and MTRR genes and serum homocysteine concentration. Reprod Biomed Online. 2016;32:407-13. PMID: 26874989 DOI: 10.1016/j.rbmo.2016.01.009

Kushnir VA, Solouki S, Sarig-Meth T, Vega MG, Albertini DF, Darmon SK, Deligdisch L, Barad DH, Gleicher N. Systemic inflammation and autoimmunity in women with chronic endometritis. Am J Reprod Immunol. 2016;75:672-7. PMID: 26952510 DOI: 10.1111/aji.12508

Lacey H, Haigh T, Westwood M, Aplin JD. Mesenchymally-derived insulin-like growth factor 1 provides a paracrine stimulus for trophoblast migration. BMC Dev Biol. 2002;2:5. PMID: 11972897 DOI: 10.1186/1471-213X-2-5

Lodigiani C, Dentali F, Banfi E, Ferrazzi P, Librè L, Quaglia I, Cafaro L, Morenghi E, Pacetti V, Zannoni E, Baggiani AM, Levi-Setti PE. The effect of parnaparin sodium on in vitro fertilization outcome: A prospective randomized controlled trial. Thromb Res. 2017;159:116-21. PMID: 28826740 DOI: 10.1016/j.thromres.2017.08.006

Mascarenhas MN, Flaxman SR, Boerma T, Vanderpoel S, Stevens GA. National, regional, and global trends in infertility prevalence since 1990: a systematic analysis of 277 health surveys. PLoS Med. 2012;9:e1001356. PMID: 23271957 DOI: 10.1371/journal.pmed.1001356

Nandi A, Bhide P, Hooper R, Gudi A, Shah A, Khan K, Homburg R. Intrauterine insemination with gonadotropin stimulation or in vitro fertilization for the treatment of unexplained subfertility: a randomized controlled trial. Fertil Steril. 2017;107:1329-35.e2. PMID: 28501361 DOI: 10.1016/j.fertnstert.2017.03.028

Nelson SM, Greer IA. The potential role of heparin in assisted conception. Hum Reprod Update. 2008;14:623-45. PMID: 18701511 DOI: 10.1093/humupd/dmn031

Ni H, Lu L, Deng J, Fan W, Li T, Yao J. Effects of Glutamate and Aspartate on Serum Antioxidative Enzyme, Sex Hormones, and Genital Inflammation in Boars Challenged with Hydrogen Peroxide. Mediators Inflamm. 2016;2016:4394695. PMID: 27777497 DOI: 10.1155/2016/4394695

Péloquin K, Brassard A, Arpin V, Sabourin S, Wright J. Whose fault is it? Blame predicting psychological adjustment and couple satisfaction in couples seeking fertility treatment. J Psychosom Obstet Gynaecol. 2018;39:64-72. PMID: 28635527 DOI: 10.1080/0167482X.2017.1289369

Practice Committee of American Society for Reproductive Medicine. Definitions of infertility and recurrent pregnancy loss: a committee opinion. Fertil Steril. 2013;99:63. PMID: 23095139 DOI: 10.1016/j.fertnstert.2012.09.023

Quaas A, Dokras A. Diagnosis and treatment of unexplained infertility. Rev Obstet Gynecol. 2008;1:69-76. PMID: 18769664

Qublan HS, Eid SS, Ababneh HA, Amarin ZO, Smadi AZ, Al-Khafaji FF, Khader YS. Acquired and inherited thrombophilia: implication in recurrent IVF and embryo transfer failure. Hum Reprod. 2006;21:2694-8. PMID: 16835215 DOI: 10.1093/humrep/del203 Ray JG, Laskin CA. Folic acid and homocyst(e)ine metabolic defects and the risk of placental abruption, pre-eclampsia and spontaneous pregnancy loss: A systematic review. Placenta. 1999;20:519-29. PMID: 10452905 DOI: 10.1053/plac.1999.0417

Ray A, Shah A, Gudi A, Homburg R. Unexplained infertility: an update and review of practice. Reprod Biomed Online. 2012;24:591-602. PMID: 22503948 DOI: 10.1016/j.rbmo.2012.02.021

Rongieres C. Assistant Reproductive Therapeutic. What therapeutic way in unexplained infertility: IUI or IVF? Med Reprod. 2015;17:100-6.

Schachter M, Raziel A, Friedler S, Strassburger D, Bern O, Ron-El R. Insulin resistance in patients with polycystic ovary syndrome is associated with elevated plasma homocysteine. Hum Reprod. 2003;18:721-7. PMID: 12660262 DOI: 10.1093/humrep/deg190

Spiridonov NA, Wong L, Zerfas PM, Starost MF, Pack SD, Paweletz CP, Johnson GR. Identification and Characterization of SSTK, a serine/threonine protein kinase essential for male fertility. Mol Cell Biol. 2005;25:4250-61. PMID: 15870294 DOI: 10.1128/MCB.25.10.4250-4261.2005

Steptoe PC, Edwards RG. Birth after the reimplantation of a human embryo. Lancet. 1978;2:366. PMID: 79723

Unni SN, Lakshman LR, Vaidyanathan K, Subhakumari KN, Menon NL. Alterations in the levels of plasma amino acids in polycystic ovary syndrome--A pilot study. Indian J Med Res. 2015;142:549-54. DOI: 10.4103/0971-5916.171281 PMID: 26658589

Weiss G, Goldsmith LT, Taylor RN, Bellet D, Taylor HS. Inflammation in reproductive disorders. Reprod Sci. 2009;16:216-29. PMID: 19208790 DOI: 10.1177/1933719108330087

Xu XY, Yang JH, Ma XM, Liu AL, Liu K, He S, Mi HY, Li L. Neonatal complications and birth defects in infants conceived by in vitro fertilization. Zhongguo Dang Dai Er Ke Za Zhi. 2015;17:350-5. PMID: 25919554 DOI: 10.7499/j.issn.1008-8830.2015.04.012

Young E. The anti-inflammatory effects of heparin and related compounds. Thrombosis Res. 2008;122:743-52. PMID: 17727922 DOI: 10.1016/j.thromres.2006.10.026

Zhong YP, Ying Y, Wu HT, Zhou CQ, Xu YW, Wang Q, Li J, Shen XT, Li J. Relationship between antithyroid antibody and pregnancy outcome following in vitro fertilization and embryo transfer. Int J Med Sci. 2012;9:121-5. PMID: 22253557 DOI: 10.7150/ijms.3467